

A Review of the Anti-inflammatory Properties of Clindamycin in the Treatment of Acne Vulgaris

James Q. Del Rosso, DO; Nicholas F. Schmidt, PhD

This article reviews anti-inflammatory properties of clindamycin, which is often used topically for the management of acne vulgaris, usually in combination with other agents. The efficacy of clindamycin in acne treatment has been shown to be sustained for more than 3 decades. It is likely that anti-inflammatory effects play an important role in the therapeutic activity of topical clindamycin.

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This article reviews anti-inflammatory properties of clindamycin, which is commonly used topically for the management of acne vulgaris. Topical clindamycin is most often utilized in combination with other topical agents, such as benzoyl peroxide, and/or a retinoid. Anti-inflammatory properties reported in association with clindamycin, expressed at the cellular and molecular level, may correlate with positive therapeutic outcomes observed during acne treatment. This article serves as a compilation of data from multiple sources that allow the reader to develop an overall appreciation for the potential therapeutic relevance of anti-inflammatory properties

associated with topical clindamycin when used to treat acne vulgaris. Given these observations, it is not surprising that the combination of clindamycin and a topical retinoid in the same topical acne treatment regimen or single vehicle formulation (ie, clindamycin phosphate 1.2%–tretinoin 0.025% gel) is known to be more therapeutically effective than either drug used alone.¹⁻³

What information is available on the pathogenesis of acne that may correlate with the clinical relevance of anti-inflammatory properties of therapeutic agents used to treat acne vulgaris?

The pathogenesis of acne is not completely understood. However, it generally is agreed that development of acne lesions occurs in relation to several factors, including hormones, primarily androgenic stimulation of the sebaceous follicle (pilosebaceous unit); excess sebum production; abnormal follicular keratinization; follicular proliferation of *Propionibacterium acnes*; stimulation of innate immune response by *P acnes*; induction and perpetuation of an inflammatory cascade by several proinflammatory enzymes, cytokines, and chemokines; and a secondary inflammatory response to follicular disruption when present.^{1,4-8}

Proinflammatory agents involved in the acne-genic inflammatory cascade include enzymes such as lipase that are known to release cytotoxic free fatty acids from sebum, as well as protease, hyaluronidase, and neuramidase. These enzymes can damage the follicular wall leading to leakage of contents into the surrounding dermis, which further propagates inflammation.⁹⁻¹⁵ Other proinflammatory agents that have been associated with

Dr. Del Rosso is from Valley Hospital Medical Center, Las Vegas, Nevada. Dr. Schmidt is an independent consultant, Roxbury, Connecticut.

Dr. Del Rosso is a consultant, researcher, and speaker for Allergan, Inc; Coria Laboratories, Ltd; Galderma Laboratories, LP; Graceway Pharmaceuticals, LLC; Intendis, Inc; Medicus Pharmaceutical Corporation; Obagi Medical Products, Inc; Onset Therapeutics; OrthoNeutrogena; Quinnova Pharmaceuticals, Inc; Ranbaxy Laboratories Ltd; SkinMedica, Inc; Stiefel Laboratories, Inc; Triax Pharmaceuticals, LLC; Unilever; and Warner Chilcott. Dr. Schmidt is a consultant for Medicus Pharmaceutical Corporation. Correspondence not available.

P. acnes include heat shock proteins, porphyrin, and squalene peroxides.^{6,13,16,17}

The pilosebaceous unit can be considered an immunocompetent organ.¹⁸ As such, it is sensitive to changes and stimuli that signal the beginning of a localized inflammatory process. Although the earliest sequence of events leading to an inflamed acne lesion remains somewhat controversial, generally it is accepted that the first objective evidence of a subclinical acne lesion is the micro-comedone.⁸ At this early stage, it is now believed that an inflammatory cascade may simultaneously occur in many cases.^{19,20} In some affected follicles but not others, progression to the development of visible comedones and/or inflammatory acne lesions occurs.⁸

Importantly, *P. acnes* releases potent low-molecular-weight (LMW) chemotactic factors and lipase enzyme during the early stages of preinflammatory and inflammatory acne lesion development.²¹⁻²⁶ These proinflammatory factors enter the dermis surrounding the follicle, thus attracting inflammatory cells such as neutrophils, monocytes/macrophages, and lymphocytes to the affected perifollicular region.^{4,14,21,27-30}

Cytokines are LMW peptides and glycoproteins that represent components of the host response providing chemical communications and transmissions between cellular constituents of the immune system. Importantly, these chemical mediators serve to direct cellular "traffic" so that an organized immunologic defense can be mounted.³¹ Within the

extensive family of biologically active cytokines that *P. acnes* can upregulate, 3 important subgroups play an important role as proinflammatory mediators during the early stages of acne lesion development: (1) chemokines, which function as chemotactic cytokines; (2) cytokines, which serve as messenger molecules; and (3) interleukins, which can serve to attract specific inflammatory cells.^{31,32} Table 1 summarizes the spectrum of proinflammatory agents and subsequent inflammatory challenges associated with *P. acnes*.

In acne lesion development, a variety of immunologically active cell types respond and contribute to the localized inflammatory cascade, including T cells, CD4⁺ cells, and CD14⁺ cells.^{18,44,47-50} Activated monocytes transform into macrophages and begin to surround and engulf foreign materials present in the perifollicular region. Intracellularly, phagocytosing neutrophils and macrophages release various degradative lysosomal enzymes and toxic chemicals such as reactive oxygen species (ROS) that attempt to remove *P. acnes* and its extracellular products.⁵¹⁻⁵⁴ With continued stimulation, the inflammatory cascade amplifies, the cycle perpetuates, and there is further damage and rupture of follicular wall integrity.^{6,55-58}

Some other proinflammatory factors have been implicated as potential pathogenic mechanisms in acne vulgaris such as granulocyte-macrophage colony-stimulating factor (GM-CSF), leukotriene B₄, and HLA-DR.^{7,14,18,47,59,60} Granulocyte-macrophage colony-stimulating factor promotes

Table 1.

Proinflammatory Agents Associated With Acne Lesion Development/ *Propionibacterium acnes*

| <i>P. acnes</i> Tissue Damage Factors | <i>P. acnes</i> Chemotactic Factors | <i>P. acnes</i> Cytokine Stimulant Factors | Proinflammatory Cytokine Early Response Factors (Chemokines/Interleukins/Interferons/Others) |
|---|---|--|--|
| Fatty acids ^{6,15,33,34} | Lipase enzyme ^{23,37} | Heat shock proteins ^{13,16-18} | IL-1 α ^{17,18,38-43} |
| Porphyrin ^{13,35,36} | Other neutrophil and immunocyte chemotactic factors ^{4,14,21,24,27-30} | | IL-1 β ^{41,44} |
| Squalene peroxides ⁶ | | | IL-6 ^{17,38,39,42,43} |
| Protease, hyaluronidase, neuramidase ^{9,11,12,14,37} | C5a ¹⁸ | | IL-8 ^{41,44,45} |
| Keratin ^{1,6} | | | TNF- α ^{17,18,41,42} |
| | | | IFN- γ ^{18,46} |

Abbreviations: TNF- α , tumor necrosis factor α ; IFN- γ , interferon- γ .

growth of leukocytes involved in the inflammatory response, and leukotriene B₄ has been shown to recruit and activate neutrophils, monocytes, and eosinophils to sites of infection. This latter cytokine is synthesized from arachidonic acid through the combined action of the 2 enzymes 5-lipoxygenase and leukotriene A₄ hydrolase.^{7,14,18,59,60} HLA-DR is a specialized protein upregulated by Langerhans cells during an infectious challenge, which helps to present antigens and activate the local immune response.⁴⁷ Complement-activated C5-derived neutrophil chemotactic factors attract more leukocytes and propagate further inflammation. Other important factors and mediators associated with *P. acnes* inflammation are IL-12, toll-like receptors, activator protein-1 transcription factor, vascular cell adhesion molecules, degranulation in macrophages, growth factors, and expression of transglutaminase and lipoxygenase.^{38,44,46,47,50,61-69}

What anti-inflammatory properties have been reported with clindamycin?

Clindamycin is a lincosamide antibiotic that exhibits an antibacterial mechanism of action by binding to the 50S ribosomal subunit of several microorganisms, including *P. acnes*. When bound to this ribosomal site, it prevents the translocation of peptidyl-transfer RNA from the A site to the P site of the ribosome during protein synthesis.^{26,70} As a result, the ribosome releases an incomplete protein, which adversely affects the viability of the organism. It also has been shown that clindamycin can interfere with the synthesis of the bacterial capsule that renders the bacterium more susceptible to phagocytosis.⁷¹

The anti-inflammatory properties of clindamycin that have been discussed in the literature are summarized in Table 2. The major indirect anti-inflammatory properties depicted in Table 2 are antibiotic activity, inhibition of protein synthesis, inhibition of lipase production, and reduction of fatty acid levels. The remaining anti-inflammatory properties described in Table 2 are referred to as direct anti-inflammatory actions of clindamycin.

What are the potential anti-inflammatory effects of clindamycin that may correlate with reduction of *P. acnes*?

Follicular proliferation of *P. acnes* generally is accepted as one of the major causative factors in acne pathogenesis.^{1,4,7} By inhibiting the growth of *P. acnes*,

clindamycin reduces the presence of many chemotactic and cytotoxic proinflammatory agents produced by the organism. As such, the antibiotic effect of clindamycin serves to downregulate the inflammatory response because the source of these inflammatory mediators, namely *P. acnes*, has been quantitatively suppressed. It has been established that clindamycin reduces the proliferation of *P. acnes*.⁷² As a result, the immunogenic potential of *P. acnes* is diminished.

How does clindamycin-associated decrease in lipase production potentially correlate with anti-inflammatory activity in acne treatment?

Follicular proliferation of *P. acnes* results in increased production of extracellular lipase, the enzyme responsible for the conversion of sebum triglycerides to free fatty acids.^{23,37} Interestingly, subinhibitory levels of clindamycin against *P. acnes* have been shown to suppress the in vitro production of lipase enzyme from 2 different strains of *Propionibacterium* species.⁷³ As a result, lower levels of free fatty acids have been observed in the sebum of patients with acne treated with clindamycin.^{15,74}

Reduced levels of follicular free fatty acids, which are believed to be comedogenic and proinflammatory, have the potential to translate into a diminished inflammatory insult for the host. Reduced levels of free fatty acids reported in association with clindamycin represent an anti-inflammatory property of this compound.^{15,73,74}

What effects have been observed with clindamycin on leukocyte chemotaxis?

Clindamycin has been shown to suppress leukocyte chemotaxis.^{23,56,75,76} Because neutrophils are recognized as an important component of acne lesion development, this activity is felt to be a clinically relevant property of clindamycin in acne treatment. How does clindamycin exert an inhibitory effect on leukocyte chemotaxis? Extracellular lipase produced by *P. acnes* has been shown to be chemotactic for neutrophils.^{23,37,72} Furthermore, *P. acnes* releases many LMW peptides that serve as potent chemotactic agents and are capable of attracting immunocytes to the infectious insult, especially neutrophils.^{4,14,21-25,27-30} As a result, the ability of clindamycin to reduce *P. acnes* organisms results in an indirect reduction of leukocyte chemotaxis. Additionally, clindamycin has been shown to markedly depress in vitro, C5-derived leukocyte chemotaxis, with minimal effect on random leukocyte migration.⁷⁵ This result was observed when clindamycin was used at concentrations less than, within, and greater than serum levels needed to

Table 2.

Anti-inflammatory Properties of Clindamycin

| Proinflammatory Factors and Components | Study Type | | Inhibits | | Enhances ^a | |
|---|--------------------------|-----------------------|--------------------------|-------------------|-----------------------|----|
| | Acne ^b | Other | Yes | No | Yes | No |
| <i>Propionibacterium acnes</i> growth | X ⁷² | | X ⁷² | | | |
| <i>P. acnes</i> protein synthesis (50S ribosomal subunit binding) | | X ^{26,70} | X ^{26,70} | | | |
| <i>P. acnes</i> lipase production | X ⁷³ | | X ⁷³ | | | |
| <i>P. acnes</i> and the release of follicular free fatty acids | X ^{15,74} | | X ^{15,74} | | | |
| Proinflammatory Chemokines (Attractants) | | | | | | |
| <i>P. acnes</i> release of leukocyte chemotactic components | X ^{23,56,75,76} | | X ^{23,56,75,76} | | | |
| IL-8 | X ^{46,c} | | | X ^{46,c} | | |
| Phagocytosis | | | | | | |
| Opsonization of bacteria for enhanced phagocytosis | | X ⁷⁷⁻⁸⁰ | | | X ⁷⁷⁻⁸⁰ | |
| Enhances and potentiates phagocytosis | | X ^{77,81,82} | | | X ^{77,81,82} | |
| Respiratory burst (ROS as O ₂ ⁻ , H ₂ O ₂) | | X ^{83,84} | X ^{83,84} | | | |
| iNOS enzymes | | X ⁸⁵ | X ⁸⁵ | | | |
| Protein kinase C enzyme/granuloma formation | X ⁸⁶ | | | X ⁸⁶ | | |
| Proinflammatory Cytokines (Primarily Monocytes) | | | | | | |
| IL-1 α | X ^{46,c} | | | X ^{46,c} | | |
| IL-1 β | X ^{46,d} | X ⁸⁷⁻⁸⁹ | X ^{46,87-89,d} | | | |
| IL-6 | X ^{46,c,e} | | X ^{46,c,e} | | | |
| IL-12p70 | X ^{46,d} | | | X ^{46,d} | | |
| IFN- γ | X ^{46,d} | | X ^{46,d} | | | |
| TNF- α | X ^{46,d} | X ^{85,87-90} | X ^{85,87-90} | X ^{46,d} | | |
| Keratinocyte Cytokines (Stimulants) | | | | | | |
| GM-CSF | X ^{46,c,e} | | X ^{46,c,e} | | | |

Abbreviations: ROS, reactive oxygen species; O₂⁻, superoxide; H₂O₂, hydrogen peroxide; iNOS, inducible nitric oxide synthase; IFN- γ , interferon- γ ; TNF- α , tumor necrosis factor α ; GM-CSF, granulocyte-macrophage colony-stimulating factor.

^aIn several instances, clindamycin can actually enhance rather than inhibit a process associated with inflammation. These enhancements can actually be beneficial therapeutically and therefore can be ranked as anti-inflammatory in nature.

^bAcne related based on available understanding of inflammatory mechanisms involved in pathogenesis.

^cFrom human keratinocytes activated by heat-killed *P. acnes*.

^dFrom human monocytes activated by heat-killed *P. acnes*.

^eInhibits at high concentration; however, the investigators suggest that the high concentration of clindamycin used "may be achievable in acne lesions after single topical application . . ."⁴⁶

exhibit antibiotic activity, which suggested a direct inhibitory effect on leukocyte chemotaxis. Furthermore, it also has been demonstrated that clindamycin can suppress leukocyte chemotaxis using *P. acnes* in vitro models.^{23,56,75,76}

What effects have been observed with clindamycin on phagocytosis and opsonization of bacteria?

When considered as an overall effect, enhanced phagocytosis may have anti-inflammatory value. Homeostatic levels of phagocytosis are considered to be an anti-inflammatory response because removal of unwanted bacteria, extracellular by-products, and other proinflammatory debris is beneficial to the host. However, if an amplified inflammatory cascade becomes chronic and excessive, continued phagocytosis can exacerbate inflammation.⁵¹⁻⁵⁴ It has been noted that macrophages can initially add to the inflammatory process; however, when activated to another bioactive form, they can actually aid in the healing process,⁹¹ which would explain how many severely inflamed sites resolve favorably without outside intervention and leave behind functionally and histologically unaffected tissues.

Clindamycin has been shown to potentiate the activity of phagocytic leukocytes.⁷⁷ Reported phagocytic activities of clindamycin include marked enhancement of phagocytosis of 4 different strains of *Bacteroides* species, elevation of the percentage of phagocytosing polymorphonuclear leukocytes obtained from gingival crevice fluids, and cellular surface changes of *Staphylococcus aureus* that render it far more susceptible to phagocytic cells.^{77,81,82} Presently it is not known if enhancement of phagocytosis by clindamycin is an operative mechanism related to the efficacy of this agent in acne treatment; however, this activity is a potential anti-inflammatory effect of clindamycin.

With regard to opsonization of bacteria, phagocytosis of *Bacteroides thetaiotaomicron* was enhanced in the presence of clindamycin.⁷⁷ At subinhibitory concentrations, clindamycin has been shown to enhance the uptake of *S. aureus* by phagocytic cells, an effect believed to be due to enhanced opsonization.⁷⁸ Additionally, the phagocytosis of *S. aureus* 502A, grown in the presence of one-third of the minimum inhibitory concentration of clindamycin, was substantially enhanced compared to the untreated control.⁷⁹ In the presence of subinhibitory concentrations of clindamycin, *Streptococcus pyogenes* organisms were denuded of cell surface M protein, thus rendering them more susceptible to phagocytosis by polymorphonuclear leukocytes.⁸⁰

What effect does clindamycin have on macrophage respiratory burst?

When macrophages digest and degrade engulfed pathogenic organisms (eg, *P. acnes*), intracellular release of cytotoxic ROS, such as superoxide (O_2^-) and hydrogen peroxide (H_2O_2), is known as respiratory or oxidative burst. Clindamycin is known to concentrate inside human phagocytic cells.^{71,83,92,93} Once inside the cell, it can potentially modulate the stimulated output of superoxides through an inhibitory effect. In vitro stimulus-activated neutrophil systems have shown that clindamycin modestly suppressed superoxide production in a formyl-methionyl-phenylalanine-stimulated system, substantially inhibited superoxide production by approximately 50% when a microbial particle-stimulated model was used, and also suppressed the release of hydrogen peroxide.⁸³ Along the same lines, clindamycin caused dose-related inhibition of in vitro superoxide formation by both untreated and pyocyanin-treated neutrophils as well as inhibition of ROS using other neutrophil-stimulated models.⁸⁴

What effect does clindamycin have on IL-1 α and IL-1 β ?

Human keratinocytes, macrophages, and monocytes are a major source of IL-1 α and IL-1 β during the cutaneous immune response to *P. acnes* proliferation, with IL-1 α expressed in correlation with comedone formation in vitro in isolated pilosebaceous units.³⁹ Furthermore, in vivo assessments of IL-1 α in comedones have shown that it can induce inflammation when released into the dermis.⁴⁰ Four in vitro studies have shown that clindamycin inhibits the production of proinflammatory IL-1 β .^{46,87-89} Inhibition of IL-1 β production may be part of the mode of action of clindamycin in the treatment of acne vulgaris.⁸⁹ Clindamycin does not inhibit the production of IL-1 α by human keratinocytes.⁴⁶

It has been shown that *P. acnes* and supernatants obtained after 72 hours of *P. acnes* growth markedly increased the induction of proinflammatory IL-1 β in human monocyte cell lines.^{32,41} An in vitro study was conducted in which human peripheral blood mononuclear cells and healthy human epidermal keratinocytes were used to measure, in part, the inhibitory effect of clindamycin on proinflammatory cytokines, including IL-1 β .⁴⁶ Heat-killed *P. acnes* served as the stimulus for upregulating cytokine production in these human cell lines. The investigators showed that clindamycin remarkably inhibited the production of proinflammatory IL-1 β by human monocytes.⁴⁶

Investigators also have demonstrated that clindamycin modulates cytokine production after exposure of mouse peritoneal macrophages to lipopolysaccharide (LPS) antigen, which is known to activate macrophages.⁸⁷ Specifically, clindamycin “decreased the intracellular expression levels of . . . interleukin 1 β (IL-1 β) and increased IL-6 expression in macrophages Our findings suggest that [clindamycin] modulates cytokine production in LPS-stimulated macrophages.”⁸⁷ In 2 other studies, clindamycin inhibited the *in vivo* production of IL-1 β in mice subjected to *Escherichia coli*-induced toxic shock endotoxin, and human acute monocytic leukemia cell line cells produced less IL-1 β when exposed to *E coli* previously treated with this drug to suppress endotoxin formation.^{88,89}

What effects have been reported with clindamycin on production of interferon- γ ?

Results showed that clindamycin inhibited the production of interferon- γ (IFN- γ) by human monocytes, which may have an effect in reducing inflammation.⁴⁶ Interferon- γ primes macrophages so that they produce degradative lysosomal activity, plays a key role in antigen processing of macrophages, and induces the production of E selectin on endothelial cells as well as intercellular adhesion molecule-1 and HLA-DR on keratinocytes; all of these processes play an important role in the attraction of leukocytes to the site of infection. Also, the generation of T cell lines from inflamed acne lesions and cytokine response after exposure to *P acnes* antigens have been explored. The results showed that the antigens were recognized and that several proinflammatory mediators were upregulated, most notably IFN- γ . The authors suggested that “IFN- γ may play a central part in the immunopathogenesis of acne.”⁹⁴ Heat-killed *P acnes* served as the stimulus for upregulating cytokine production in human peripheral blood mononuclear cells, with clindamycin shown to substantially inhibit the production of IFN- γ by human monocytes.⁴⁶

What effects have been reported with clindamycin on production of tumor necrosis factor α ?

Tumor necrosis factor α (TNF- α) is a multifunctional cytokine that plays an important role in immunologic response, including upregulation of many prostaglandins, collagenase enzymes, and various inflammatory cells, as well as release of several cytokines, such as IL-1, IL-6, IL-8, and GM-CSF.^{41,42,46} One of the earliest expressions of the inflammatory response is the activation of cellular adhesion molecules that serve to traffic inflammatory cells into peripheral tissues,

a response that appears to be under the control of TNF- α and IL-8. Tumor necrosis factor α can be produced by human keratinocytes when stimulated with UV light or LPS endotoxins that activate macrophages.⁴² Overall, TNF- α can upregulate and induce a broad range of secondary proinflammatory effects in response to pathogenic organisms, including *P acnes*.

Multiple studies that employed various *in vitro* models demonstrated that clindamycin can inhibit the production of TNF- α , including LPS-induced macrophages. This inhibitory effect could potentially translate into therapeutic anti-inflammatory activity at the clinical level.^{85,87-90} One study reported that clindamycin did not inhibit the production of TNF- α when *P acnes* was used to stimulate human peripheral monocytes.⁴⁶

What effect does clindamycin have on nitric oxide synthase enzymes and nitric oxide formation?

Nitric oxide (NO) is synthesized from arginine and oxygen by various inducible nitric oxide synthase (iNOS) enzymes. Nitric oxide is an important cellular mediator of inflammatory and vascular responses in various organ systems, including skin. Some proinflammatory cytokines can induce excessive quantities of NO via induction of iNOS, ultimately stimulating inflammatory responses that can lead to cellular damage.⁹³ In one study, clindamycin was shown to decrease the ability of group B streptococci to stimulate iNOS accumulation in macrophages, an effect that results in decreased production of NO.⁸⁵

What effects have been reported with clindamycin on production of IL-6?

IL-6 is an immune cytokine with a wide range of proinflammatory and anti-inflammatory properties. In the presence of IL-1, IL-6 is secreted by macrophages, T lymphocytes, and keratinocytes, with decreased IL-6 production resulting in a marked decline in the acute inflammatory response. IL-6, a proinflammatory mediator in inflammatory acne, has been shown to be secreted by peritoneal macrophages using an *in vitro P acnes*-induced cytokine production model.⁴⁴

Healthy human keratinocyte cells were used in an *in vitro* study to evaluate the inhibitory effect of clindamycin on IL-6 induction when heat-killed *P acnes* served as the stimulus for upregulation.⁴⁶ The study showed that use of clindamycin at the highest concentration (ie, 30 $\mu\text{g/mL}$) inhibited the production of IL-6 by human keratinocytes. The authors also pointed out that the inhibitory tissue concentration is achievable after topical application of clindamycin.⁴⁶

What effects have been reported with clindamycin on production of GM-CSF?

Granulocyte-macrophage colony-stimulating factor is a proinflammatory proteinaceous cytokine secreted by macrophages, other immune cells, and keratinocytes. It is a component of the inflammatory cascade that can stimulate production of large numbers of macrophages in response to an inflammatory challenge.⁴⁶

The same in vitro study described earlier for IL-6 also was employed to evaluate the inhibitory effect of clindamycin on GM-CSF production by human keratinocytes. Heat-killed *P acnes* served as the stimulus for upregulating the GM-CSF cytokine. Once again, the investigators demonstrated that clindamycin decreased production of GM-CSF by healthy human keratinocytes.⁴⁶

What other effects or lack of effects have been reported with clindamycin that relate to anti-inflammatory properties?

Clindamycin did not inhibit induction of IL-1 α and IL-8 in human keratinocytes when heat-killed *P acnes* was used as the activating agent nor did it inhibit the production of IL-12p70 in human monocytes activated by heat-killed *P acnes*.⁴⁶ It also has been observed that clindamycin does not have an inhibitory effect on the activity of protein kinase C, an agent whose presence has been linked to more severe forms of acne.⁸⁶

What conclusions can be drawn on anti-inflammatory effects of clindamycin from the information described?

A myriad of articles have been published, as referenced here, discussing several immunomodulatory properties of clindamycin. Many of these anti-inflammatory effects may correlate with therapeutic activity in acne vulgaris, especially those effects related to inhibition of cytokines that promote the innate inflammatory response.^{39,40,46,87-89} It should be noted that anti-inflammatory properties of clindamycin often are referred to as indirect and direct. The term *indirect* should not be confused with lesser activity. It is not entirely clear which anti-inflammatory properties play major or minor roles in acne treatment; however, the net effect of these anti-inflammatory properties suggest that the efficacy of clindamycin in acne treatment is not likely related to antibacterial activity alone. As the efficacy of clindamycin in acne treatment has been shown to be sustained for more than 3 decades, despite the emergence of clindamycin-resistant *P acnes* strains in some patients, it is likely that anti-inflammatory effects

play an important role in the therapeutic activity of topical clindamycin.⁹⁵

The clinical ramifications suggested by these in vitro observations are supported by the results of many clinical studies that demonstrate the efficacy of topical clindamycin in acne treatment.⁹⁵ It also has been shown that topical clindamycin alone or in combination with topical tretinoin is able to reduce both inflammatory and noninflammatory acne lesions.^{2,95-98} Importantly, it is not suggested that topical clindamycin be used as monotherapy for the treatment of acne; however, monotherapy studies do substantiate its therapeutic value in acne treatment.^{1,95-99}

REFERENCES

1. Berson DS, Shalita AR. The treatment of acne: the role of combination therapies. *J Am Acad Dermatol.* 1995;32(5, pt 3):S31-S41.
2. Zouboulis CC, Derumeaux L, Decroix J, et al. A multicentre, single-blind, randomized comparison of a fixed clindamycin phosphate/tretinoin gel formulation applied once daily and a clindamycin formulation applied twice daily in the topical treatment of acne vulgaris. *Br J Dermatol.* 2000;143:498-505.
3. Richter JR, Förström LR, Kiistala UO, et al. Efficacy of the fixed 1.2% clindamycin phosphate, 0.025% tretinoin gel formulation (Velac) and a proprietary 0.025% tretinoin gel formulation (Aberela) in the topical control of facial acne. *J Eur Acad Dermatol Venereol.* 1998;11:227-233.
4. Gollnick HP, Zouboulis CC, Akamatsu H, et al. Pathogenesis and pathogenesis related treatment of acne. *J Dermatol.* 1991;18:489-499.
5. Cunliffe WJ. The sebaceous gland and acne—40 years on. *Dermatology.* 1998;196:9-15.
6. Toyoda M, Morohashi M. Pathogenesis of acne. *Med Electron Microsc.* 2001;34:29-40.
7. Zouboulis CC, Piquero-Martin J. Update and future of systemic acne treatment. *Dermatology.* 2003;206:37-53.
8. Jappe U. Pathological mechanisms of acne with special emphasis on *Propionibacterium acnes* and related therapy. *Acta Derm Venereol.* 2003;83:241-248.
9. Hoeffler U. Enzymatic and hemolytic properties of *Propionibacterium acnes* and related bacteria. *J Clin Microbiol.* 1977;6:555-558.
10. Van Vlem B, Vanholder R, De Paepe P, et al. Immunomodulating effects of antibiotics: literature review. *Infection.* 1996;24:275-291.
11. Ingham E, Holland KT, Gowland G, et al. Purification and partial characterization of an acid phosphatase (EC 3.1.3.2) produced by *Propionibacterium acnes*. *J Gen Microbiol.* 1980;118:59-65.
12. Puhvel S, Reisner RM. The production of hyaluronidase (hyaluronate lyase) by *Corynebacterium acnes*. *J Invest Dermatol.* 1972;58:66-70.

13. Holland KT, Aldana O, Bojar RA, et al. *Propionibacterium acnes* and acne. *Dermatology*. 1998;196:67-68.
14. Burkhart CG, Burkhart CN, Lehmann PF. Acne: a review of immunologic and microbiologic factors. *Postgrad Med J*. 1999;75:328-331.
15. Thomsen RJ, Stranieri A, Knutson D. Topical clindamycin treatment of acne. *Arch Dermatol*. 1980;116:1031-1034.
16. Farrar MD, Ingham E, Holland KT. Heat shock proteins and inflammatory acne vulgaris: molecular cloning, over-expression and purification of a *Propionibacterium acnes* GroEL and DnaK homologue. *FEMS Microbiol Lett*. 2000;191:183-186.
17. Graham GM, Farrar MD, Cruse-Sawyer JE, et al. Pro-inflammatory cytokine production by human keratinocytes stimulated with *Propionibacterium acnes* and *P acnes* GroEL. *Br J Dermatol*. 2004;150:421-428.
18. Koreck A, Pivarcsi A, Dobozy A, et al. The role of innate immunity in the pathogenesis of acne. *Dermatology*. 2003;206:96-105.
19. Bhambri S, Del Rosso JQ, Bhambri A. Pathogenesis of acne vulgaris: recent advances. *J Drugs Dermatol*. 2009;7:615-618.
20. Harper JC, Thiboutot DM. Pathogenesis of acne: recent research advances. *Adv Dermatol*. 2003;19:1-10.
21. Puhvel SM, Sakamoto M. The chemoattractant properties of comedonal components. *J Invest Dermatol*. 1978;71:324-329.
22. Puhvel SM, Sakamoto M. Cytotoxin production by comedonal bacteria. *J Invest Dermatol*. 1980;74:36-40.
23. Lee WL, Shalita AR, Sunthralingam K, et al. Neutrophil chemotaxis to *Propionibacterium acnes* lipase and its inhibition. *Infect Immun*. 1982;35:71-78.
24. Webster GF, Leyden JJ. Characterization of serum-independent polymorphonuclear leukocyte chemotactic factors produced by *Propionibacterium acnes*. *Inflammation*. 1980;4:261-271.
25. Webster GF. Inflammation in acne vulgaris. *J Am Acad Dermatol*. 1995;33(2, pt 1):247-253.
26. Douthwaite S. Interaction of the antibiotics clindamycin and lincomycin with *Escherichia coli* 23S ribosomal RNA. *Nucleic Acids Res*. 1992;20:4717-4720.
27. Webster GF, Leyden JJ, McGinley KJ, et al. Suppression of polymorphonuclear leukocyte chemotactic factor production in *Propionibacterium acnes* by subliminal inhibitory concentrations of tetracycline, ampicillin, minocycline and erythromycin. *Antimicrob Agents Chemother*. 1982;21:770-772.
28. Brown SK, Shalita AR. Acne vulgaris. *Lancet*. 1998;351:1871-1876.
29. Zouboulis CC. Acne: current aspects on pathology and treatment. *Dermatol Experiences*. 1999;1:6-37.
30. Piquero-Martin J. *Acne Manejo Racional*. 3rd ed. Caracas, Venezuela: Corpografica; 2000.
31. Stienhoff M, Luger TA. The skin cytokine network. In: Bos JD, ed. *Skin Immune System: Cutaneous Immunology and Clinical Immunodermatology*. 3rd ed. Boca Raton, FL: CRC Press; 2005:349-372.
32. Pastore S, Cavani A, Albanesi C, et al. Chemokines of human skin. In: Bos JD, ed. *Skin Immune System: Cutaneous Immunology and Clinical Immunodermatology*. 3rd ed. Boca Raton, FL: CRC Press; 2005:373-392.
33. Zouboulis CC. Is acne vulgaris a genuine inflammatory disease? *Dermatology*. 2001;203:277-279.
34. Akamatsu H, Tomita T, Horio T. Effects of Roxithromycin on the production of lipase and neutrophil chemotactic factor by *Propionibacterium acnes*. *Dermatology*. 2002;204:277-280.
35. Gribbon EM, Shoemith JG, Cunliffe WJ, et al. The microaerophily and photosensitivity of *Propionibacterium acnes*. *J Appl Bacteriol*. 1994;77:583-590.
36. Kjeldstad B, Johnsson A, Sandberg S, et al. Influence of pH on porphyrin production in *Propionibacterium acnes*. *Arch Dermatol Res*. 1984;276:396-400.
37. Holland KT. Nutrition of cutaneous resident microorganisms. In: Noble WC, ed. *The Skin Microflora and Microbial Skin Disease*. Cambridge, England: Cambridge University Press; 1993:33-72.
38. Luger TA, Schwarz T. Evidence for an epidermal cytokine network. *J Invest Dermatol*. 1990;95:1005-1011.
39. Ansel J, Perry P, Brown J, et al. Cytokine modulation of keratinocyte cytokines. *J Invest Dermatol*. 1990;94(suppl 6):101S-107S.
40. Ingham E, Eady EA, Goodwin CE, et al. Pro-inflammatory levels of interleukin-1 α -like bioactivity are present in the majority of open comedones in acne vulgaris. *J Invest Dermatol*. 1992;98:895-901.
41. Vowels BR, Yang S, Leyden JJ. Induction of proinflammatory cytokines by a soluble factor of *Propionibacterium acnes*: implications for chronic inflammatory acne. *Infect Immun*. 1995;63:3158-3165.
42. Kock A, Schwarz T, Kirnbauer R, et al. Human keratinocytes are a source for tumor necrosis factor α : evidence for synthesis and release upon stimulation with endotoxin or ultraviolet light. *J Exp Med*. 1990;172:1609-1614.
43. Kupper TS. The activated keratinocyte: a model for inducible cytokine production by non-bone marrow-derived cells in cutaneous inflammatory and immune responses. *J Invest Dermatol*. 1990;94(suppl 6):146S-150S.
44. Kim J, Ochoa M, Krutzyk S, et al. Activation of toll-like receptor 2 in acne triggers inflammatory cytokine responses. *J Immunol*. 2002;169:1535-1541.
45. Chen Q, Koga T, Uchi H, et al. *Propionibacterium acnes*-induced IL-8 production may be mediated by NF- κ B activation in human monocytes. *J Dermatol Sci*. 2002;29:97-103.
46. Kuwahara K, Kitazawa T, Kitagaki H, et al. Nadifloxacin, an antiacne quinolone antimicrobial, inhibits the

- production of proinflammatory cytokines by human peripheral mononuclear cells and normal human keratinocytes. *J Dermatol Sci*. 2005;38:47-55.
47. Jappe U, Ingham E, Henwood J, et al. *Propionibacterium acnes* and inflammation in acne; *P acnes* has T-cell mitogenic activity. *Br J Dermatol*. 2002;146:202-209.
 48. Jeremy AH, Holland DB, Roberts SG, et al. Inflammatory events are involved in acne lesion initiation. *J Invest Dermatol*. 2003;121:20-27.
 49. Norris JFB, Cunliffe WJ. A histological and immunological study of early acne lesions. *Br J Dermatol*. 1988;118:651-659.
 50. Liu PT, Krutzok SR, Kim J, et al. Cutting edge: all-*trans*-retinoic acid downregulates TLR2 expression and function. *J Immunol*. 2005;174:2467-2470.
 51. Webster GF, Leyden JJ, Tsai CC, et al. Polymorphonuclear leukocyte lysosomal release in response to *Propionibacterium acnes* in vitro and its enhancement by sera from patients with inflammatory acne. *J Invest Dermatol*. 1980;74:398-401.
 52. Webster GF, Kligman AM. A method for the assay of inflammatory acne of inflammatory mediators in follicular casts. *J Invest Dermatol*. 1979;73:266-268.
 53. Miyachi Y, Yoshioka A, Imamura S, et al. Effect of antibiotics on the generation of reactive oxygen species. *J Invest Dermatol*. 1986;86:449-453.
 54. Akamatsu H, Komura J, Miyachi Y, et al. Suppressive effects of linoleic acid on neutrophil oxygen metabolism and phagocytosis. *J Invest Dermatol*. 1990;95:271-274.
 55. Webster GF, McGinley KJ, Leyden JJ. Inhibition of lipase production in *Propionibacterium acnes* by sub-minimal-inhibitory concentrations of tetracycline and erythromycin. *Br J Dermatol*. 1981;104:453-457.
 56. Akamatsu H, Kurokawa I, Nishijima S, et al. Inhibition of neutrophil chemotactic factor production in comedonal bacteria by subminimal inhibitory concentrations of erythromycin. *Dermatology*. 1992;185:41-43.
 57. Camisa C, Eisenstat B, Ragaz A, et al. The effects of retinoids on neutrophil function in vitro. *J Am Acad Dermatol*. 1982;6(4, pt 2)(suppl):620-629.
 58. Akamatsu H, Horio T. The possible role of reactive oxygen species generated by neutrophils in mediating acne inflammation. *Dermatology*. 1998;196:82-85.
 59. Zouboulis CC. Exploration of retinoid activity and the role of inflammation in acne: issues affecting the future directions for acne therapy. *J Eur Acad Dermatol Venereol*. 2001;15(suppl 3):63-67.
 60. Bray MA. Retinoids are potent inhibitors of the generation of rat leukocyte leukotriene B₄-like activity in vitro. *Eur J Pharmacol*. 1984;98:61-67.
 61. Shalita AR, Wei-Li L. Inflammatory acne. *Dermatol Clin*. 1983;1:361-364.
 62. Massey A, Mowbray JF, Noble WC. Complement activation by *Corynebacterium acnes*. *Br J Dermatol*. 1978;98:583-584.
 63. Webster GF, Leyden JJ, Norman ME, et al. Complement activation in acne vulgaris: in vitro studies with *Propionibacterium acnes* and *Propionibacterium granulosum*. *Infect Immun*. 1978;22:523-529.
 64. Burkhart CG, Cantrill J, Butcher CL, et al. *Propionibacterium acnes*: interaction with complement and development of an enzyme-linked immunoassay for the detection of antibody. *Int J Dermatol*. 1999;38:200-203.
 65. Millikan LE. The rationale for using a topical retinoid for inflammatory acne. *Am J Clin Dermatol*. 2003;4:75-80.
 66. Czernielewski J, Michel S, Bouclier M, et al. Adapalene biochemistry and the evolution of a new topical retinoid for treatment of acne. *J Eur Acad Dermatol Venereol*. 2001;15(suppl 3):5-12.
 67. Gille J, Paxton LL, Lawley TJ, et al. Retinoic acid inhibits the regulated expression of vascular cell adhesion molecule-1 by cultured dermal microvascular endothelial cells. *J Clin Invest*. 1997;99:492-500.
 68. Fumarulo R, Conese M, Riccardi S, et al. Retinoids inhibit the respiratory burst and degranulation of stimulated human polymorphonuclear leukocytes. *Agents Actions*. 1991;34:339-344.
 69. Shroot B, Michel S. Pharmacology and chemistry of adapalene. *J Am Acad Dermatol*. 1997;36(6, pt 2):S96-S103.
 70. Vasquez D. *Inhibitors of Protein Biosynthesis*. Berlin, Germany: Springer-Verlag; 1979.
 71. Gemmell CG. Interactions between clindamycin and immune function. *Rev Contemp Pharmacother*. 1992;3:321-327.
 72. Tan H. Topical antibacterial treatments for acne vulgaris. *Am J Clin Dermatol*. 2004;5:79-84.
 73. Unkles SE, Gemmell CG. Effect of clindamycin, erythromycin, lincomycin, and tetracycline on growth and extracellular lipase production by propionibacteria in vitro. *Antimicrob Agents Chemother*. 1982;21:39-43.
 74. Pablo GM, Fulton JE Jr. Sebum: analysis by infrared spectroscopy. II. the suppression of free fatty acids by systemically administered antibiotics. *Arch Dermatol*. 1975;111:734-735.
 75. Esterly NB, Furey NL, Flanagan LE. The effect of antimicrobial agents on leukocyte chemotaxis. *J Invest Dermatol*. 1978;70:51-55.
 76. Akamatsu H, Nishijima S, Takahashi M, et al. Effects of subminimal concentrations of erythromycin, tetracycline, clindamycin, and minocycline on the neutrophil chemotactic factor production in *Propionibacterium acnes* biotypes 1-5. *J Dermatol*. 1991;18:247-251.
 77. Veringa EM, Lambe DS Jr, Ferguson DA Jr, et al. Enhancement of opsonophagocytosis of *Bacteroides* spp. by clindamycin in subinhibitory concentrations. *J Antimicrob Chemother*. 1989;23:577-587.

78. Veringa EM, Verhoef J. Influence of subinhibitory concentrations of clindamycin on opsonophagocytosis of *Staphylococcus aureus*, a protein-A-dependent process. *Antimicrob Agents Chemother*. 1986;30:796-797.
79. Milatovic D, Braveny I, Verhoef J. Clindamycin enhances opsonization of *Staphylococcus aureus*. *Antimicrob Agents Chemother*. 1983;24:413-417.
80. Gemmell CG, Peterson PK, Schmeling D, et al. Potentiation of opsonization and phagocytosis of *Streptococcus pyogenes* following growth in the presence of clindamycin. *J Clin Invest*. 1981;67:1249-1256.
81. Eick S, Pfister W, Fiedler D, et al. Clindamycin promotes phagocytosis and intracellular killing of periodontopathogenic bacteria by crevicular granulocytes: an in vitro study. *J Antimicrob Chemother*. 2000;46:583-588.
82. Mascellino MT, De Vito ML, Maclean Feeney E, et al. Phagocytosis and killing of A-protein positive *Staphylococcus aureus* in the presence of low doses of antibiotics. *Drugs Exp Clin Res*. 1989;15:63-69.
83. Hand WL, Hand DL, King-Thompson NL. Antibiotic inhibition of the respiratory burst response in human polymorphonuclear leukocytes. *Antimicrob Agents Chemother*. 1990;34:863-870.
84. Ras GJ, Anderson R, Taylor GW, et al. Clindamycin, erythromycin, and roxithromycin inhibit the proinflammatory interactions of *Pseudomonas aeruginosa* pigments with human neutrophils in vitro. *Antimicrob Agents Chemother*. 1992;36:1236-1240.
85. Brinkmann KC, Talati AJ, Akbari RE, et al. Group B streptococci exposed to rifampin or clindamycin (versus ampicillin or cefotaxime) stimulate reduced production of inflammatory mediators by murine macrophages. *Pediatr Res*. 2005;57:419-423.
86. Webster GF, Toso SM, Hegemann L. Inhibition of a model of in vitro granuloma formation by tetracyclines and ciprofloxacin. *Arch Dermatol*. 1994;130:748-752.
87. Nakano T, Hiramatsu K, Kishi K, et al. Clindamycin modulates inflammatory-cytokine induction in lipopolysaccharide-stimulated mouse peritoneal macrophages. *Antimicrob Agents Chemother*. 2003;47:363-367.
88. Hirata N, Hiramatsu K, Hishi K, et al. Pretreatment of mice with clindamycin improves survival of endotoxic shock by modulating the release of inflammatory cytokines. *Antimicrob Agents Chemother*. 2001;45:2638-2642.
89. Kishi K, Hirai K, Hiramatsu K, et al. Clindamycin suppresses endotoxin released by ceftazidime-treated *Escherichia coli* O55:B5 and subsequent production of tumor necrosis factor- α and interleukin 1 β . *Antimicrob Agents Chemother*. 1999;43:616-622.
90. Stevens DL, Bryant AE, Hackett SP. Antibiotic effects on bacterial viability, toxin production, and host response. *Clin Infect Dis*. 1995;20(suppl 2):S154-S157.
91. Duffield JS. The inflammatory macrophage: a story of Jekyll and Hyde. *Clin Sci (Lond)*. 2003;104:27-38.
92. Hand WL, Hand DL. Influence of pentoxifylline and its derivatives on antibiotic uptake and superoxide generation by human phagocytic cells. *Antimicrob Agents Chemother*. 1995;39:1574-1579.
93. Dell'Anna ML, Camera E, Picardo M. Free radicals. In: Bos JD, ed. *Skin Immune System: Cutaneous Immunology and Clinical Immunodermatology*. 3rd ed. Boca Raton, FL: CRC Press; 2005:287-313.
94. Mouser PE, Baker BS, Seaton ED, et al. *Propionibacterium acnes*-reactive T helper-1 cells in the skin of patients with acne vulgaris [letter]. *J Invest Dermatol*. 2003;121:1226-1228.
95. Simonart T, Dramaix M. Treatment of acne with topical antibiotics: lessons learned from clinical studies. *Br J Dermatol*. 2005;153:395-403.
96. Schlessinger J, Menter A, Gold M, et al. Clinical safety and efficacy studies of a novel formulation combining 1.2% clindamycin phosphate and 0.025% tretinoin for the treatment of acne vulgaris. *J Drugs Dermatol*. 2007;6:607-615.
97. Rosen T, Waisman M. Topically administered clindamycin in the treatment of acne vulgaris and other dermatologic disorders. *Pharmacotherapy*. 1981;1:201-205.
98. Shalita AR, Myers JA, Krochmal L, et al. The safety and efficacy of clindamycin phosphate foam 1% versus clindamycin phosphate topical gel 1% for the treatment of acne vulgaris. *J Drugs Dermatol*. 2005;4:48-56.
99. Gollnick H, Cunliffe W, Berson D, et al; Global Alliance to Improve Outcomes in Acne. Management of acne: a report from the Global Alliance to Improve Outcomes in Acne. *J Am Acad Dermatol*. 2003;49(suppl 1):S1-S37.