

Actinic Keratosis as a Marker of Field Cancerization in Excision Specimens of Cutaneous Malignancies

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PRACTICE POINTS

- Clinically apparent and subclinical actinic keratoses usually are present in patients, a concept known as field cancerization, and it is important to treat both types of lesions.
- Actinic keratoses are present in the field of cutaneous malignancies, including basal cell carcinoma, squamous cell carcinoma, and melanoma.

Field cancerization is the process in which a singular cell accumulates genetic mutations following carcinogen exposure and then divides to create a “field” of monoclonal premalignant cells. In this study, microscopically identified actinic keratoses (AKs) were used as markers of field cancerization in all excision specimens of squamous cell carcinomas (SCCs), basal cell carcinomas (BCCs), and malignant melanomas (MMs) received by our institution’s dermatopathology department over a 3- to 6-month period. Our findings provide additional evidence for the theory of field cancerization, its association with cutaneous malignancies, and the need to assess the extent of field damage when determining treatment strategies.

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The concept of field cancerization was first proposed in 1953 by Slaughter et al¹ in their study of oral squamous carcinomas. Their findings of multifocal patches of premalignant disease, abnormal tissue surrounding tumors, multiple localized primary tumors, and tumor recurrence following surgical resection was suggestive of a field of dysplastic cells with malignant potential.¹ Since then, modern molecular techniques have been used to establish a genetic basis for this model in many different types of cancer, including cutaneous malignancies.²⁻⁴ The field begins from a singular stem cell, which undergoes one or more genetic changes that allow for a growth advantage compared to surrounding cells. The stem cell then divides, forming a patch of clonal daughter cells that displace the surrounding normal epithelium. Growth of this patch eventually leads to a dysplastic field of monoclonal cells, which notably does not yet show invasive growth or metastatic behavior. Over time, continued carcinogenic exposure results in additional genetic alterations among different cells in the field, which leads to new subclonal proliferations that share common clonal origin but exhibit unique genetic changes. Eventually, transformative events may occur, resulting in cells with invasive and metastatic properties, thus forming a carcinoma.⁵

In the case of cutaneous malignancies, UV radiation in the form of UVA and UVB rays is the most

common source of carcinogenesis. It is well established that UV radiation has numerous effects on the body, including but not limited to local and systemic immunosuppression, alteration of signal transduction pathways, and the development of mutations in DNA via direct damage by UVB or indirect damage by free radical formation with UVA.^{6,7} Normally, DNA is protected from UV radiation-induced genetic alteration by the p53 gene, *TP53*. As such, damage to this gene is highly associated with cancer induction. One study found that more than 90% of squamous cell carcinomas (SCCs) and more than 50% of basal cell carcinomas (BCCs) contain UV-like mutations in *TP53*.⁸ The concept of field cancerization suggests that because the skin surrounding cutaneous malignancies has been exposed to the same chronic UV light as the initial lesion, it is at an increased risk for genetic abnormalities and thus possible malignant transformation.

Actinic keratoses (AKs) are common neoplasms of the skin that generally are regarded as precancerous lesions or may be considered to be the earliest stage of SCC *in situ*.⁹ Actinic keratoses usually develop as a consequence of chronic exposure to UV radiation and often are clinically apparent as erythematous scaly papules in sun-exposed areas (Figure 1).¹⁰ They also are identified histologically as atypical keratinocytes along the basal layer of the epidermis with possible enlargement, hyperchromatic nuclei, lack of maturation, mitotic figures, inflammatory infiltrate, and/or hyperkeratosis.¹⁰ Furthermore, the genetic changes associated with AKs are well documented and are strongly associated with changes to p53.¹¹ Given these characteristics, AKs serve as good markers of genetic damage with potential for malignancy. In this study, we used histologically identified AKs to assess the presence of field damage in the tissue immediately surrounding excision specimens of SCCs, BCCs, and malignant melanomas (MMs).

Methods

This study was approved by the Program for the Protection of Human Subjects at the Icahn School of Medicine at Mount Sinai (New York, New York) prior to initiation. All cutaneous specimens submitted to the dermatopathology service for consultation between April 2013 and June 2013 were reviewed for inclusion in this study. Data collection was extended for MMs to include all specimens from January 2013 to June 2013 given the limited number of cases in the original data collection period.

Initial screening for this study was done electronically and assessed for a diagnosis of SCC (Figure 2),

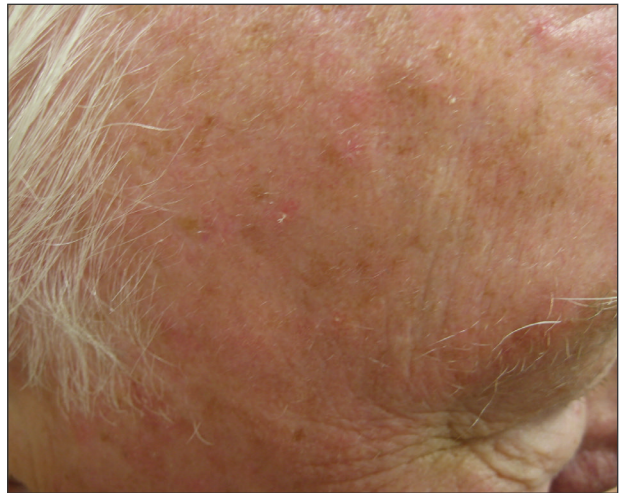


Figure 1. Scaly erythematous papules typical of actinic keratosis. Copyright Gary Goldenberg, MD.

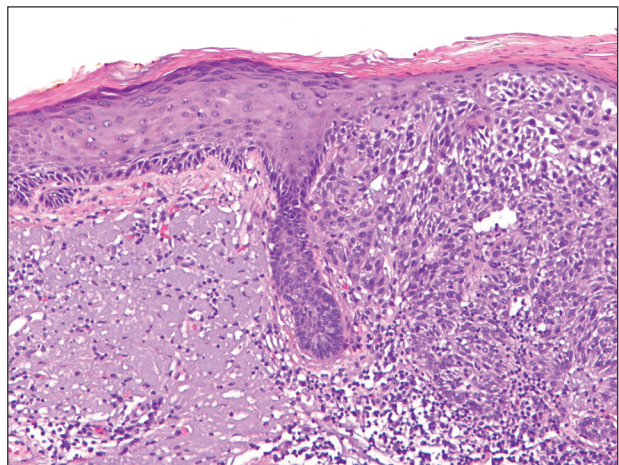


Figure 2. On the left, an actinic keratosis demonstrates atypical keratinocytes along the basal layer with hyperchromatic nuclei and atypical maturation. Separated by a section of histologically normal epithelium, a squamous cell carcinoma is seen on the right (H&E, original magnification $\times 200$).

BCC (Figure 3), or MM (Figure 4) as determined by a board-certified dermatopathologist (G.G.). The resulting pool of specimens was then screened to include only excision specimens and to exclude curettage specimens and superficial specimens that lacked dermis. In this study, we chose to look at reexcisions rather than initial biopsies so that there was a greater likelihood of having an intact epidermis surrounding a malignancy that could be assessed for the presence of AKs as markers for field cancerization. Specimens were examined in full via serial transverse cross-sections at 3-mm intervals.

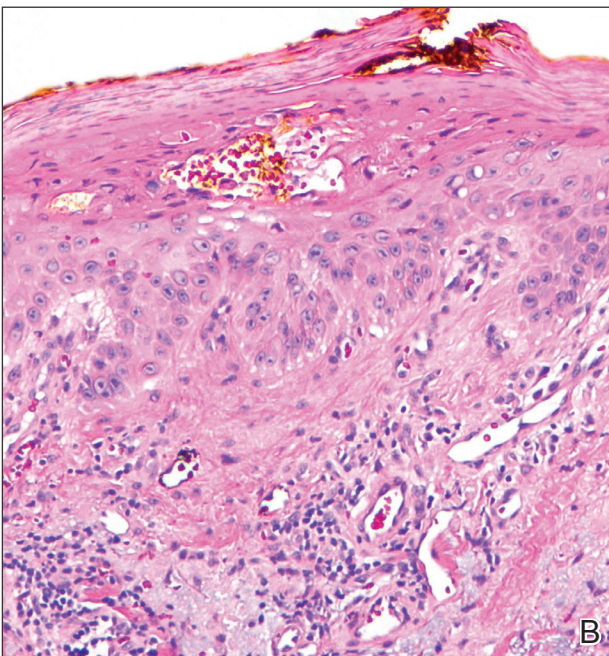
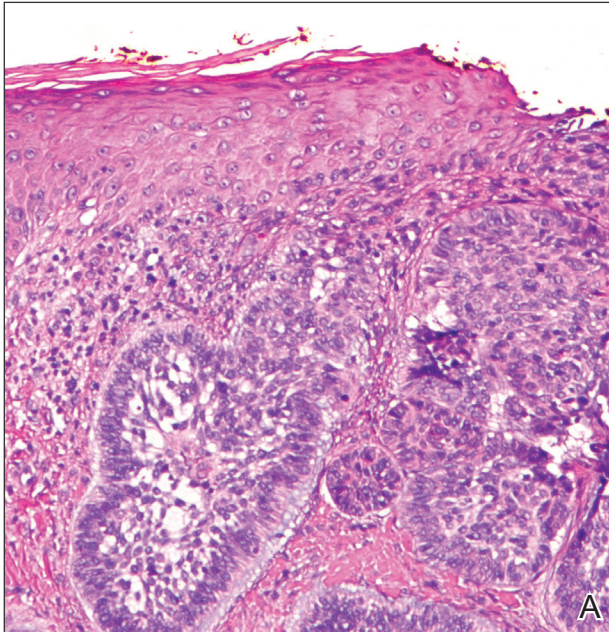


Figure 3. Residual basal cell carcinoma (A)(H&E, original magnification $\times 200$). Actinic keratosis from the same excision with notable parakeratosis and solar elastosis in the dermis (B)(H&E, original magnification $\times 200$).

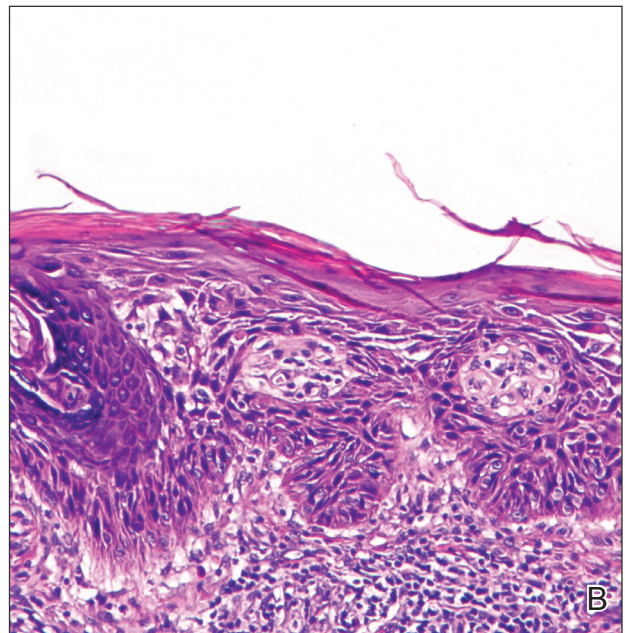
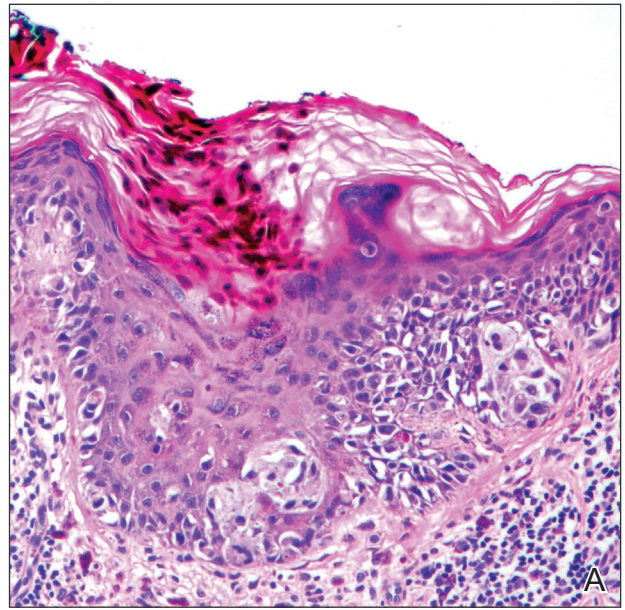


Figure 4. Malignant melanoma (A)(H&E, original magnification $\times 200$). Incidental actinic keratosis in the same excision specimen, both images exhibit a lymphocytic infiltrate.

Additional step sections were obtained at smaller intervals when margins were close or unclear.

Selected cases were reassessed by a board-certified dermatopathologist (G.G.) to confirm the diagnosis and to assess for the presence of at least 1 AK within the specimen sample that

was separated from the original malignancy by histologically normal-appearing cells. Samples were also assessed for the presence of an AK within 0.1 mm of the distal lateral margins of the tissue sample. Information regarding patient age, gender, lesion location, lesion type, and specimen size was

collected for each sample. In accordance with institutional review board protocol, research data were collected without any protected health information. All analyses and results were deidentified and stored on a password-protected computer database. Statistical analysis was performed using SPSS software. When applicable, $P < .05$ was considered to indicate statistical significance.

Results

There were 205 cases that passed the initial screening filters, of which 56 were excluded due to the presence of curettage or lack of a sufficient tissue sample. Of the remaining 149 cases, the distribution by malignancy type was tabulated along with the percentage of observed AKs. If an AK was observed, the percentage that had an AK at the lateral margins (marginal AK) was determined (Table 1). A χ^2 analysis determined that AKs were observed significantly more often in SCC specimens (57% [35/61]) than BCC (33% [21/64]) or malignant melanoma (25% [6/24]) specimens ($P = .0125$).

Statistics regarding patient age and gender as well as specimen size were stratified by malignancy type (Table 2). Using a receiver operating characteristic curve and the Youden index, an optimal cutoff of older than 67 years was determined to increase probability of observing an AK ($P = .077$) with sensitivity of 0.531 and specificity of 0.529. The distribution of specimen excision location for each malignancy type is shown in Table 3.

A multivariate analysis was performed to determine if the variables of patient age, gender, biopsy size, malignancy type (SCC, BCC, or MM), or cancer location (head, neck, trunk, arms, or legs) were independently useful in predicting whether an AK would be observed in the excision specimen. The significance of variables in the logistic regression model was assessed using a backward stepwise regression selection procedure entering variables if $P < .15$ and excluding variables if $P > .25$. Significant variables in predicting the occurrence of AK were SCC malignancy type ($P = .007$; odds ratio [OR], 2.61) and location on the head ($P = .044$; OR, 2.39) and arms ($P = .042$; OR, 2.55).

Comment

The χ^2 analysis of our data showed that SCC specimens were significantly more likely to have an associated AK than either BCCs or MMs ($P = .0125$), which is not surprising given that AKs are considered by many to be early-stage SCCs.¹² It is important to note, however, that BCCs and MMs both had non-negligible rates of associated AKs. Although BCC and MM do not arise from the same background of

Table 1.

Incidence Rates of AK by Type of Cutaneous Malignancy

	Malignancy Type			Total
	BCC	SCC	MM	
No. of specimens examined, n (%)	64 (43)	61 (41)	24 (16)	149 (100)
No. of specimens with observed AKs, n (%) ^a	21 (33)	35 (57)	6 (25)	62 (42)
No. of specimens with marginal AKs (%) ^b	13 (62)	20 (57)	2 (33)	35 (56)

Abbreviations: AK, actinic keratosis; BCC, basal cell carcinoma; SCC, squamous cell carcinoma; MM, malignant melanoma.

^aPercentage represents specimens by malignancy type.

^bPercentage represents observed AKs by malignancy type.

Table 2.

Study Population and Specimen Statistics

Characteristic	Malignancy Type			Total
	BCC	SCC	MM	
Mean age (SD), y	72 (13)	63 (13)	75 (13)	68 (14)
Sex, %				
Male	53	57	58	56
Female	47	43	42	44
Mean specimen size (SD), cm ²	2.7 (2.2)	2.1 (2.3)	6.2 (5.5)	3.0 (3.3)

Abbreviations: BCC, basal cell carcinoma; SCC, squamous cell carcinoma; MM, malignant melanoma.

genetic changes as SCC, this finding is noteworthy because it demonstrates definitive field damage with malignant potential in the area surrounding these cutaneous malignancies.

Our data also showed that there was a significantly greater association of AKs in malignancies located on the head ($P = .044$) and arms ($P = .042$), possibly because these 2 areas tend to be the most sun

Table 3.

Distribution of Specimen Excision Location

Excision Site	Malignancy Type			Total (N=149)
	BCC (n=64)	SCC (n=61)	MM (n=24)	
Head, n (%)	12 (18.8)	11 (18.0)	9 (37.5)	32 (21.5)
Neck, n (%)	1 (1.6)	1 (1.6)	1 (4.2)	3 (2.0)
Trunk, n (%)	37 (57.8)	20 (32.8)	10 (41.7)	67 (45.0)
Arms, n (%)	8 (12.5)	17 (27.9)	3 (12.5)	28 (18.8)
Legs, n (%)	6 (9.4)	12 (19.7)	1 (4.2)	19 (12.8)

Abbreviations: BCC, basal cell carcinoma; SCC, squamous cell carcinoma; MM, malignant melanoma.

exposed and thus are more likely to have sustained field damage as evidenced by the higher percentage of AKs. A study by Jonason et al¹³ described a similar finding in which sun-exposed skin exhibited significantly more frequent ($P=.04$) and larger ($P=.02$) clonal patches of mutated p53 keratinocytes than sun-protected skin.

It is likely that the field damage surrounding the cutaneous lesions in our study is actually greater than what we reported because the AK was present at the margin of the excision specimens the majority of the time (56%), which suggests that there likely may have been more AKs found if a wider area surrounding the malignancy had been studied given that AKs often are at the periphery of the lesion and may be missed by a small excision. Fewer marginal AKs were observed with MM cases, possibly because the excision specimens were more than double the size of SCC or BCC excisions. Furthermore, there likely is to be more damage than what can be appreciated by visual changes alone.

Kanjilal et al¹⁴ used polymerase chain reaction and DNA sequencing to demonstrate numerous p53 mutations in nonmalignant-appearing skin surrounding BCCs and SCCs. Brennan et al¹⁵ found p53 mutations in surgical margins of excised SCCs considered to be tumor free by histopathologic analysis in more than half of the specimens studied. Notably, tumor recurrence was significantly more likely in areas where mutations were found and no tumor recurrence was seen in areas free of p53 mutations ($P=.02$).¹⁵ Tabor et al⁴ similarly found genetically altered fields in histologically clear surgical margins of SCCs but also showed that local tumor recurrence following excision had more molecular markers in common with the nonresected premalignant field than it did with the primary

tumor. Thus, these studies provide a genetic basis for field damage that can exist even in histologically benign-appearing cells.

We believe our findings are clinically relevant, as they provide additional evidence for the theory of field cancerization as demonstrated by the non-negligible rates of AKs and thus field damage with malignant potential in the skin immediately surrounding cutaneous malignancies. The limitations of our study, however, include a small sample size; no consideration of the effects of prior topical, field, or systemic treatments; and lack of a control group. Nevertheless, our findings emphasize the importance of assessing the extent of field damage when determining treatment strategies. Clinicians treating cutaneous malignancies should consider the need for field therapy, especially in sun-exposed regions, to avoid additional primary tumors.¹⁶ Further research is needed, however, to identify optimal methods for quantifying field damage clinically and determining the most effective treatment strategies.

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