

KARL B. SHPARGEL
Department of Neurosciences,
Cleveland Clinic, Cleveland, OH

WALID JALABI, PhD
Department of Neurosciences,
Cleveland Clinic, Cleveland, OH

YONGMING JIN
Department of Neurosciences,
Cleveland Clinic, Cleveland, OH

ALISHER DADABAYEV, MD
Department of Stem Cell Biology and
Regenerative Medicine, Cleveland Clinic,
Cleveland, OH

MARC S. PENN, MD, PhD
Departments of Cardiovascular Medicine,
Biomedical Engineering, and Stem Cell
Biology and Regenerative Medicine; and
Director, Bakken Heart-Brain Institute,
Cleveland Clinic, Cleveland, OH

BRUCE D. TRAPP, PhD
Chairman, Department of Neurosciences,
Cleveland Clinic, Cleveland, OH

Preconditioning paradigms and pathways in the brain

■ ABSTRACT

Preconditioning is a phenomenon in which the brain protects itself against future injury by adapting to low doses of noxious insults. Preconditioning stimuli include ischemia, low doses of endotoxin, hypoxia, hypothermia and hyperthermia, cortical spreading depression, anesthetics, and 3-nitropropionic acid, among others. Understanding of the mechanisms underlying preconditioning has been elusive, but NMDA receptor activation, nitric oxide, inflammatory cytokines, and suppression of the innate immune system appear to have a role. Elucidation of the endogenous cell survival pathways involved in preconditioning has significant clinical implications for preventing neuronal damage in susceptible patients.

The brain relies upon internal defense mechanisms for protection from injurious stimuli. Preconditioning is a phenomenon whereby low doses of these noxious insults shield the brain from future insults rather than inflicting damage. Preconditioning stimuli include but are not limited to transient global and focal ischemia,¹⁻⁴ cortical spreading depression,⁵⁻⁷ brief episodes of seizure, exposure to anesthetic inhalants,⁸⁻¹⁰ low doses of endotoxin (lipopolysaccharide [LPS]),^{11,12} hypothermia and hyperthermia,^{13,14} and 3-nitropropionic acid treatment.^{15,16}

Depending on the specific preconditioning stimulus, a state of neuronal tolerance can be established in at least two temporal profiles: one in which the trigger induces protection within minutes (rapid or acute tolerance),¹⁷ and one in which the protected state develops after a delay of several hours to days (delayed tolerance).⁴ Some preconditioning paradigms induce both

phases of ischemic tolerance, while others can induce only the acute phase or only the delayed phase.¹⁸⁻²¹ The acute phase is most likely due to rapid posttranslational modifications of proteins.^{22,23} In contrast, the delayed phase is dependent on de novo protein synthesis.^{24,25}

Preconditioning by ischemic tolerance was first identified in the heart by Murry et al,²⁶ and was subsequently found to occur in the brain^{4,27} and a variety of organs including the liver, intestine, kidney, and lung. Preconditioning stimuli can be cross-tolerant, safeguarding against other types of injury. For example, endotoxin preconditioning can protect against subsequent ischemia and vice versa. Thus, there may be some overlapping mechanisms in preconditioning, and unraveling these pathways may uncover an arsenal of neuroprotective therapeutic targets. In this review, we will compare different preconditioning paradigms and discuss potential mechanisms in initiating brain ischemic tolerance.

■ PARADIGMS TO ESTABLISH PRECONDITIONING

Refinement of various preconditioning models is of great clinical significance. Cardiovascular or cerebrovascular surgery has a negative impact on brain function due to stoppage of blood flow during surgery. In fact, more than 25% of patients who receive coronary artery bypass surgery suffer from temporary or permanent memory loss.^{28,29} As a result, it is of premier importance to develop strategies to protect the brain either prior to vascular surgeries or in patients at high risk of stroke. While it would be dangerous and impractical to precondition at-risk patients with ischemia, the identification of underlying preconditioning mechanisms may lead to safer therapeutic factors that can be administered before surgery.

Ischemia

Global ischemic preconditioning in the brain is accomplished by occlusion of the bilateral common carotid

All authors reported that they have no financial relationships that pose a potential conflict of interest with this article.

arteries. In contrast, in focal ischemic preconditioning, occlusion of one side of the middle cerebral artery is induced for about 1 to 20 minutes, depending on methods and animal species.^{4,30-32} Twenty-four hours after ischemic preconditioning, stroke is induced in these animals. Preconditioning-induced neuroprotection is observed not only in terms of infarct volume but also in terms of neurological scores and behavior studies.

Lipopolysaccharide

Tolerance to ischemic injury can also be induced by a small dose of LPS injected into the peritoneal cavity. Dosages vary from 0.05 to 1 mg/kg body weight in small rodents such as mice and rats.^{11,33-36} This dose of LPS usually does not bring abnormal signs and symptoms to the animals. The ischemic protection yields a reduction of infarct volume of approximately 30%. This tolerant state can be sustained for about 1 week, with maximum protection occurring around 2 to 3 days after injection of LPS.

Hypoxia

A relatively convenient method for preconditioning animals is hypoxic exposure. Animals are put in a chamber in which oxygen and nitrogen proportions can be controlled. Oxygen concentration usually ranges from 8% to 13% with normobaric pressure. Exposure time ranges from 1 to 6 hours. Twenty-four to 72 hours later, transient or permanent focal stroke is induced in the animals.³⁷⁻⁴⁰ Hypoxia-preconditioned neuroprotection usually starts at 1 to 3 days with a significant reduction of infarct size. Hypoxic preconditioning has also been demonstrated for in vitro neuron culture models using oxygen-glucose deprivation injury.⁴¹

3-Nitropropionic acid

3-Nitropropionic acid (3-NP) is an irreversible inhibitor of succinate dehydrogenase, an enzyme required for oxidative phosphorylation and adenosine triphosphate production. When applied at low doses 1 to 4 days before ischemia, 3-NP can lead to ischemic tolerance in the forebrain of gerbils and rats.^{16,42,43} The dose ranges from 1 to 20 mg/kg body weight.¹⁶ Such treatment significantly improves neurological behavior and increases neuronal survival in the CA1 region of hippocampus. In addition, 3-NP preconditioning induces tolerance to hypoxia in hippocampal slice preparations.^{15,44}

Hypothermia and hyperthermia

Hypothermia is a well-characterized protective procedure used during and after cerebral surgery. It is also reported that brief hypothermic or hyperthermic exposure can also lead to ischemic tolerance. The tempera-

tures adopted range from 25°C to 32°C^{13,45,46} in hypothermia and from 42°C to 43°C in hyperthermia.¹⁴

Cortical spreading depression

Cortical spreading depression is defined as the electrophysiologic phenomenon of slowly propagating transient depolarization waves across the cortex. Usually 5 M of potassium chloride is infused into the cortex, or a cotton pad soaked with the solution is put on the surface of dura mater, which results in depolarization, firing of neurons, and cortical spreading depression. Cortical spreading depression induces a prolonged phase of ischemic tolerance that lasts 1 to 7 days.^{5,6,47,48}

Anesthetics

Exposure to volatile anesthetics such as isoflurane and halothane within pharmacologic concentration ranges also confers delayed-phase ischemic tolerance of the brain.^{8-10,49}

■ MOLECULAR PRECONDITIONING PATHWAYS

Mechanistically, cellular preconditioning can be subdivided into intrinsic neuronal pathways (preventing excitotoxic damage, signaling through anti-apoptotic molecules, and treatment by neurotrophic factors) or extrinsic nonneuronal pathways (peripheral cytokine production, microglial activation, and regulation of the cerebrovascular system). Several neuroprotective molecules are expressed and signal through multiple cell types both within and peripheral to the brain, so that assigning an exact source and paradigm for preconditioning pathways has proven difficult.

NMDA receptor activation and excitotoxicity protection

In neurons, ischemic tolerance is mediated largely by the activation of the *N*-methyl-D-aspartate (NMDA) glutamate receptors through increases in intracellular calcium.⁵⁰⁻⁵² Although glutamate receptor activation is generally believed to be responsible for much of the neuronal damage caused by excitotoxicity, it appears to also be implicated in the establishment of preconditioning. One study demonstrated that exposure of cortical cell cultures to low levels of glutamate activated NMDA receptors in preconditioning.⁵⁰ In addition, preconditioning by oxygen-glucose deprivation was blocked when an NMDA antagonist was applied. NMDA receptor activation can induce a tolerant state through rapid adaptation of the voltage-dependent calcium flux. In addition, activation of NMDA receptors leads to rapid release of brain-derived neurotrophic factor, which then binds to and activates its cognate receptor, receptor tyrosine kinase B. Both NMDA and tyrosine kinase B receptors activate nuclear factor- κ B (NF κ B), a transcription factor involved

in protecting neurons against insults. In sublethal ischemic preconditioning, activation of NF κ B and its translocation from the cytosol to the nucleus was required for the development of late cerebral protection against severe ischemia or epilepsy.⁵³ Other key mediators involved in synaptic NMDA receptor–dependent neuroprotection are phosphatidylinositol 3-kinase (PI3K), Akt, and glycogen synthase kinase 3- β .⁵⁴

Preconditioning with cortical spreading depression results in the downregulation of the excitatory amino acid transporters EAAT1 and EAAT2 from cerebral cortex plasma membranes.⁵⁵ Although these transporters are normally involved in glutamate uptake, it has been suggested that the influx of sodium that occurs during excitotoxicity may cause their reversal and result in additional glutamate release. Downregulating these transporters may thus contribute to ischemic tolerance.

Nitric oxide

Nitric oxide (NO) may play a key role as a mediator of the neuronal ischemic preconditioning response, either in conjunction with or independent of NMDA receptor activation. Both the inhibition of nitric oxide synthase (NOS) and the scavenging of NO during preconditioning significantly attenuated the induced neuronal tolerance, and neither endothelial NOS nor neuronal NOS knockout mice showed protection from rapid ischemic preconditioning.^{56,57} Treatment with the inducible NOS (iNOS) inhibitor aminoguanidine abolished the induced protection. The mechanisms responsible for NO-induced tolerance are not clear. Downregulation of the glutamate transporter GLT-1 might play a role.⁵⁸ A common link to NMDA receptor activation and NO is p21^{ras} (Ras). Preconditioning induces p21^{ras} activation in an NMDA- and NO-dependent manner and leads to the downstream activation of Raf kinase, mitogen-activated protein kinases, and extracellular regulated kinase.⁵⁹ Inhibition of these kinases attenuates subsequent protection from ischemia.^{60,61} Pharmacologic inhibition of Ras, as well as a dominant negative Ras mutant, blocked preconditioning, whereas a constitutively active form of Ras promoted neuroprotection against lethal insults. An important consideration regarding NO is also that preconditioning by volatile anesthetics appears to involve NO pathways.⁹

NO and reactive oxygen species (ROS) are also implicated in regulating the peripheral cerebrovascular system. Ischemia generated by occlusion of the middle cerebral artery causes defects in cerebrovascular function for not only the infarcted area but also the surrounding ischemic region. LPS preconditioning has been reported in some cases to increase this

regional cerebral blood flow both before and after ischemia.^{1,21,36,62–64} LPS also improves microvascular perfusion.^{33,64} It was recently reported that LPS-stimulated cerebral blood flow is induced through reactive oxygen and nitrogen species (ROS or NO).¹ Mouse knockouts of iNOS (NO production) or of the nox2 subunit of NADPH oxidase (ROS production) eliminated the LPS-upregulated cerebrovascular activity. Furthermore, blockage of these ROS and NO pathways reduced the preconditioning effect of LPS. Therefore, LPS may play a more direct role in preventing ischemic damage by increasing blood availability to the affected brain region.

Inflammatory cytokines and the innate immune system

LPS, a component of the gram-negative bacterial cell wall, can illicit a potent innate immune response. While this systemic inflammatory response can be destructive (at doses of 5 mg/kg),⁶⁵ tolerable LPS doses of 0.05 to 1 mg/kg injected intraperitoneally render the brain,¹¹ heart,^{66,67} liver,^{68,69} kidneys,⁷⁰ and pancreas⁷¹ transiently resistant to subsequent ischemic injury. This preconditioning paradigm relies on the ability of a peripheral signal to cross into multiple organ systems. LPS injected into the gut can signal through peritoneal macrophages and circulating monocytes. Toll-like receptor 4 is a pattern-recognition receptor that binds to pathogen-associated molecular patterns in LPS and initiates a signaling cascade through the NF κ B pathway. This pathway culminates in the expression and secretion of several proinflammatory cytokines to fight off the infection and anti-inflammatory cytokines to control the immune response.

The major output of LPS signaling is innate production of proinflammatory cytokines to fight infection and clear cellular debris. Central cytokines, including tumor necrosis factor– α (TNF α), interleukin-6 (IL-6), and interleukin-1 beta (IL-1 β), can be neurodestructive if administered after ischemia. TNF α administration by cerebroventricular injection after ischemia augmented the extent of injury, and blockage of TNF α signaling proved neuroprotective.^{11,72,73} However, in LPS preconditioning, cytokine production precedes ischemia. Intracisternal injections of TNF α before middle cerebral artery occlusion (MCAO) were protective in reducing the infarct size of pretreated mice.⁷⁴ Furthermore, intracisternal injection of ceramide analog, a downstream component of the TNF α signaling pathway, was also capable of reducing the MCAO infarct area.⁷⁵ Preischemic treatment with IL-6 and IL-1 also reduced neuronal damage.^{76,77} TNF α knockout mice eliminated the LPS

protective phenotype,⁷² demonstrating that cytokine production is a critical feature of LPS preconditioning in ischemia. Additionally, ischemic damage in the absence of LPS preconditioning was exacerbated in TNF α receptor 1 knockout mice.^{78,79} Consistently, TNF α protein levels are upregulated after LPS treatment but are downregulated following LPS-preconditioned MCAO.⁷² A unifying theme in LPS preconditioning comprises early activation of the innate immune system with ensuing suppression in ischemia.

As a potential mechanism, the initial inflammatory response induced by LPS appears to render the innate immune system hyporesponsive to subsequent insults such as ischemia. This may occur by persistence of anti-inflammatory cytokines produced by the primary insult. These molecules are expressed in tandem with proinflammatory cytokines to control the innate immune response, but may also play a role in delayed preconditioning. For instance, intravenous or intracerebroventricular IL-10 injection can reduce the infarct size with MCAO.⁸⁰ Alternatively, several proinflammatory cytokine signaling pathways may be downregulated by negative feedback inhibition.^{20,81} This inhibition may occur extracellularly, using soluble cytokine receptors, decoy receptors, or receptor antagonists. For example, intravenous injection of IL-1 receptor antagonist can provide neuroprotection against ischemic injury from MCAO.^{82,83} Cytokine feedback inhibitors that act intracellularly are also induced with the innate immune response. Intracellular inhibition may involve direct downregulation of cytokine transcription (peroxisome proliferator-activated receptor gamma [PPAR- γ]) or inhibition of intracellular signaling pathways that promote cytokine production (suppressor of cytokine signaling [SOCS] and PI3K). Antisense mRNA knockdown of SOCS-3 exacerbates ischemic injury from MCAO.⁸⁴ The MCAO infarction area is increased after treatment with PPAR- γ antagonists and decreased by PPAR- γ agonists.^{85,86} Administration of compounds that increase PI3K signaling is also capable of reducing ischemic damage.⁸⁷ Thus, several defense mechanisms designed to suppress the innate immune response may play an active role in LPS ischemic preconditioning.

Role of microglia in ischemic preconditioning

Microglia represent the resident central nervous system (CNS) component of the innate immune system. Microglia and macrophages become activated with ischemia in the infarcted and surrounded area.⁸⁸ Upon activation in ischemia, microglia will become phagocytic and secrete a multitude of noxious chemokines and cytokines.⁸⁹ Accordingly, anti-inflammatory anti-

biotics such as doxycycline and minocycline reduce microglial activation and diminish the ischemic infarction area.⁹⁰ Preconditioning the brain with LPS ameliorates microglial activation, neutrophil infiltration, and circulating monocyte activation following MCAO.³⁵ However, primary ischemic damage is not correlated with CNS infiltration of peripheral leukocytes but rather with an increase in proliferating resident microglial cells.⁹¹ Alternatively, microglia can exhibit neuroprotective properties within the brain.⁹² In fact, greater ischemic damage from longer periods of MCAO is correlated with fewer proliferating microglia, suggesting a protective microglial role.⁹¹ Consistently, ablation of proliferating microglia increases the infarction area following MCAO.⁹³ Therefore, microglia can be protective in ischemia, and preconditioning with LPS may render microglia more capable of reacting to ischemic conditions.

CONCLUSIONS

Preconditioning represents an adaptive response to prime the brain for protection against future injury. Elucidation of these endogenous cell survival pathways has significant clinical implications for preventing neuronal damage in susceptible patients. For this reason, understanding the underlying mechanisms in establishing a tolerant state will be a critical step in adapting preconditioning for safe patient applications. The field of ischemic research has made great strides in deciphering causative preconditioning factors but has been hampered by the complex, multifactorial nature of preconditioning paradigms. The study of tolerance is further complicated by the fact that signaling takes place both peripheral to and within the brain in multiple cell types. Future research will require the exploration of interactions between multiple pathways and roles of individual cell types in establishing ischemic tolerance. Only with a more thorough understanding of preconditioning mechanisms can we adapt these pathways for the most efficient and protective treatments.

REFERENCES

1. Kunz A, Park L, Abe T, et al. Neurovascular protection by ischemic tolerance: role of nitric oxide and reactive oxygen species. *J Neurosci* 2007; 27:7083–7093.
2. Simon RP, Niuro M, Gwinn R. Prior ischemic stress protects against experimental stroke. *Neurosci Lett* 1993; 163:135–137.
3. Chen J, Simon R. Ischemic tolerance in the brain. *Neurology* 1997; 48:306–311.
4. Kitagawa K, Matsumoto M, Tagaya M, et al. 'Ischemic tolerance' phenomenon found in the brain. *Brain Res* 1990; 528:21–24.
5. Kawahara N, Ruetzler CA, Klatzo I. Protective effect of spreading depression against neuronal damage following cardiac arrest cerebral ischaemia. *Neurol Res* 1995; 17:9–16.
6. Taga K, Patel PM, Drummond JC, Cole DJ, Kelly PJ. Transient

- neuronal depolarization induces tolerance to subsequent forebrain ischemia in rats. *Anesthesiology* 1997; 87:918–925.
7. Kobayashi S, Harris VA, Welsh FA. Spreading depression induces tolerance of cortical neurons to ischemia in rat brain. *J Cereb Blood Flow Metab* 1995; 15:721–727.
 8. Baughman VL, Hoffman WE, Miletich DJ, Albrecht RE, Thomas C. Neurologic outcome in rats following incomplete cerebral ischemia during halothane, isoflurane, or N₂O. *Anesthesiology* 1988; 69:192–198.
 9. Kapinya KJ, Lowl D, Futterer C, et al. Tolerance against ischemic neuronal injury can be induced by volatile anesthetics and is inducible NO synthase dependent. *Stroke* 2002; 33:1889–1898.
 10. Zheng S, Zuo Z. Isoflurane preconditioning induces neuroprotection against ischemia via activation of P38 mitogen-activated protein kinases. *Mol Pharmacol* 2004; 65:1172–1180.
 11. Tasaki K, Ruetzler CA, Ohtsuki T, et al. Lipopolysaccharide pre-treatment induces resistance against subsequent focal cerebral ischemic damage in spontaneously hypertensive rats. *Brain Res* 1997; 748:267–270.
 12. Zimmermann C, Ginis I, Furuya K, et al. Lipopolysaccharide-induced ischemic tolerance is associated with increased levels of ceramide in brain and in plasma. *Brain Res* 2001; 895:59–65.
 13. Nishio S, Yunoki M, Chen ZF, Anzivino MJ, Lee KS. Ischemic tolerance in the rat neocortex following hypothermic preconditioning. *J Neurosurg* 2000; 93:845–851.
 14. Ota A, Ikeda T, Xia XY, Xia YX, Ikenoue T. Hypoxic-ischemic tolerance induced by hyperthermic pretreatment in newborn rats. *J Soc Gynecol Investig* 2000; 7:102–105.
 15. Riepe MW, Niemi WN, Megow D, Ludolph AC, Carpenter DO. Mitochondrial oxidation in rat hippocampus can be preconditioned by selective chemical inhibition of succinic dehydrogenase. *Exp Neurol* 1996; 138:15–21.
 16. Sugino T, Nozaki K, Takagi Y, Hashimoto N. 3-Nitropropionic acid induces ischemic tolerance in gerbil hippocampus in vivo. *Neurosci Lett* 1999; 259:9–12.
 17. Perez-Pinzon MA, Xu GP, Dietrich WD, Rosenthal M, Sick TJ. Rapid preconditioning protects rats against ischemic neuronal damage after 3 but not 7 days of reperfusion following global cerebral ischemia. *J Cereb Blood Flow Metab* 1997; 17:175–182.
 18. Stagliano NE, Perez-Pinzon MA, Moskowitz MA, Huang PL. Focal ischemic preconditioning induces rapid tolerance to middle cerebral artery occlusion in mice. *J Cereb Blood Flow Metab* 1999; 19:757–761.
 19. Perez-Pinzon MA, Born JG. Rapid preconditioning neuroprotection following anoxia in hippocampal slices: role of the K⁺ ATP channel and protein kinase C. *Neuroscience* 1999; 89:453–459.
 20. Kariko K, Weissman D, Welsh FA. Inhibition of toll-like receptor and cytokine signaling—a unifying theme in ischemic tolerance. *J Cereb Blood Flow Metab* 2004; 24:1288–1304.
 21. Hoyte LC, Papadakis M, Barber PA, Buchan AM. Improved regional cerebral blood flow is important for the protection seen in a mouse model of late phase ischemic preconditioning. *Brain Res* 2006; 1121:231–237.
 22. Nakase H, Heimann A, Uranishi R, Riepe MW, Kempinski O. Early-onset tolerance in rat global cerebral ischemia induced by a mitochondrial inhibitor. *Neurosci Lett* 2000; 290:105–108.
 23. Meller R, Cameron JA, Torrey DJ, et al. Rapid degradation of Bim by the ubiquitin-proteasome pathway mediates short-term ischemic tolerance in cultured neurons. *J Biol Chem* 2006; 281:7429–7436.
 24. Kirino T. Ischemic tolerance. *J Cereb Blood Flow Metab* 2002; 22:1283–1296.
 25. Stenzel-Poore MP, Stevens SL, Xiong Z, et al. Effect of ischaemic preconditioning on genomic response to cerebral ischaemia: similarity to neuroprotective strategies in hibernation and hypoxia-tolerant states. *Lancet* 2003; 362:1028–1037.
 26. Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* 1986; 74:1124–1136.
 27. Kitagawa K, Matsumoto M, Kuwabara K, et al. 'Ischemic tolerance' phenomenon detected in various brain regions. *Brain Res* 1991; 561:203–211.
 28. Roach GW, Kanchuger M, Mangano CM, et al. Adverse cerebral outcomes after coronary bypass surgery. Multicenter Study of Perioperative Ischemia Research Group and the Ischemia Research and Education Foundation Investigators. *N Engl J Med* 1996; 335:1857–1863.
 29. Newman MF, Kirchner JL, Phillips-Bute B, et al. Longitudinal assessment of neurocognitive function after coronary-artery bypass surgery. *N Engl J Med* 2001; 344:395–402.
 30. Corbett D, Crooks P. Ischemic preconditioning: a long term survival study using behavioural and histological endpoints. *Brain Res* 1997; 760:129–136.
 31. Barone FC, White RE, Spera PA, et al. Ischemic preconditioning and brain tolerance: temporal histological and functional outcomes, protein synthesis requirement, and interleukin-1 receptor antagonist and early gene expression. *Stroke* 1998; 29:1937–1950.
 32. Wu C, Zhan RZ, Qi S, et al. A forebrain ischemic preconditioning model established in C57Black/Crj6 mice. *J Neurosci Methods* 2001; 107:101–106.
 33. Dawson DA, Furuya K, Gotoh J, et al. Cerebrovascular hemodynamics and ischemic tolerance: lipopolysaccharide-induced resistance to focal cerebral ischemia is not due to changes in severity of the initial ischemic insult, but is associated with preservation of microvascular perfusion. *J Cereb Blood Flow Metab* 1999; 19:616–623.
 34. Ahmed SH, He YY, Nassief A, et al. Effects of lipopolysaccharide priming on acute ischemic brain injury. *Stroke* 2000; 31:193–199.
 35. Rosenzweig HL, Lessov NS, Henshall DC, et al. Endotoxin preconditioning prevents cellular inflammatory response during ischemic neuroprotection in mice. *Stroke* 2004; 35:2576–2581.
 36. Furuya K, Zhu L, Kawahara N, et al. Differences in infarct evolution between lipopolysaccharide-induced tolerant and nontolerant conditions to focal cerebral ischemia. *J Neurosurg* 2005; 103:715–723.
 37. Bernaudin M, Nedelec AS, Divoux D, et al. Normobaric hypoxia induces tolerance to focal permanent cerebral ischemia in association with an increased expression of hypoxia-inducible factor-1 and its target genes, erythropoietin and VEGF, in the adult mouse brain. *J Cereb Blood Flow Metab* 2002; 22:393–403.
 38. Kulinskii VI, Minakina LN, Gavrilina TV. Neuroprotective effect of hypoxic preconditioning: phenomenon and mechanisms. *Bull Exp Biol Med* 2002; 133:202–204.
 39. Miller BA, Perez RS, Shah AR, et al. Cerebral protection by hypoxic preconditioning in a murine model of focal ischemia-reperfusion. *Neuroreport* 2001; 12:1663–1669.
 40. Prass K, Scharff A, Ruscher K, et al. Hypoxia-induced stroke tolerance in the mouse is mediated by erythropoietin. *Stroke* 2003; 34:1981–1986.
 41. Liu J, Ginis I, Spatz M, Hallenbeck JM. Hypoxic preconditioning protects cultured neurons against hypoxic stress via TNF-alpha and ceramide. *Am J Physiol Cell Physiol* 2000; 278:C144–C153.
 42. Kuroiwa T, Yamada I, Endo S, Hakamata Y, Ito U. 3-Nitropropionic acid preconditioning ameliorates delayed neurological deterioration and infarction after transient focal cerebral ischemia in gerbils. *Neurosci Lett* 2000; 283:145–148.
 43. Horiguchi T, Kis B, Rajapakse N, Shimizu K, Busija DW. Opening of mitochondrial ATP-sensitive potassium channels is a trigger of 3-nitropropionic acid-induced tolerance to transient focal cerebral ischemia in rats. *Stroke* 2003; 34:1015–1020.
 44. Aketa S, Nakase H, Kamada Y, Hiramatsu K, Sakaki T. Chemical preconditioning with 3-nitropropionic acid in gerbil hippocampal slices: therapeutic window and the participation of adenosine receptor. *Exp Neurol* 2000; 166:385–391.
 45. Nishio S, Chen ZF, Yunoki M, et al. Hypothermia-induced ischemic tolerance. *Ann N Y Acad Sci* 1999; 890:26–41.
 46. Yunoki M, Nishio S, Ukita N, Anzivino MJ, Lee KS. Hypothermic preconditioning induces rapid tolerance to focal ischemic injury in the rat. *Exp Neurol* 2003; 181:291–300.
 47. Kawahara N, Ruetzler CA, Mies G, Klatzo I. Cortical spreading depression increases protein synthesis and upregulates basic fibroblast growth factor. *Exp Neurol* 1999; 158:27–36.
 48. Yanamoto H, Hashimoto N, Nagata I, Kikuchi H. Infarct tolerance against temporary focal ischemia following spreading depression in rat brain. *Brain Res* 1998; 784:239–249.
 49. McAuliffe JJ, Joseph B, Vorhees CV. Isoflurane-delayed preconditioning reduces immediate mortality and improves striatal function in adult mice after neonatal hypoxia-ischemia. *Anesth Analg* 2007;

- 104:1066–1077.
50. Grabb MC, Choi DW. Ischemic tolerance in murine cortical cell culture: critical role for NMDA receptors. *J Neurosci* 1999; 19:1657–1662.
 51. Kato H, Liu Y, Araki T, Kogure K. MK-801, but not anisomycin, inhibits the induction of tolerance to ischemia in the gerbil hippocampus. *Neurosci Lett* 1992; 139:118–121.
 52. Kasischke K, Ludolph AC, Riepe MW. NMDA-antagonists reverse increased hypoxic tolerance by preceding chemical hypoxia. *Neurosci Lett* 1996; 214:175–178.
 53. Blondeau N, Widmann C, Lazdunski M, Heurteaux C. Activation of the nuclear factor- κ B is a key event in brain tolerance. *J Neurosci* 2001; 21:4668–4677.
 54. Soriano FX, Papadia S, Hofmann F, et al. Preconditioning doses of NMDA promote neuroprotection by enhancing neuronal excitability. *J Neurosci* 2006; 26:4509–4518.
 55. Douen AG, Akiyama K, Hogan MJ, et al. Preconditioning with cortical spreading depression decreases intras ischemic cerebral glutamate levels and down-regulates excitatory amino acid transporters EAAT1 and EAAT2 from rat cerebral cortex plasma membranes. *J Neurochem* 2000; 75:812–818.
 56. Atochin DN, Clark J, Demchenko IT, Moskowitz MA, Huang PL. Rapid cerebral ischemic preconditioning in mice deficient in endothelial and neuronal nitric oxide synthases. *Stroke* 2003; 34:1299–1303.
 57. Cho S, Park EM, Zhou P, et al. Obligatory role of inducible nitric oxide synthase in ischemic preconditioning. *J Cereb Blood Flow Metab* 2005; 25:493–501.
 58. Yamada T, Kawahara K, Kosugi T, Tanaka M. Nitric oxide produced during sublethal ischemia is crucial for the preconditioning-induced down-regulation of glutamate transporter GLT-1 in neuron/astrocyte co-cultures. *Neurochem Res* 2006; 31:49–56.
 59. Gonzalez-Zulueta M, Feldman AB, Klesse LJ, et al. Requirement for nitric oxide activation of p21^{ras}/extracellular regulated kinase in neuronal ischemic preconditioning. *Proc Natl Acad Sci U S A* 2000; 97:436–441.
 60. Kurkinen K, Keinanen R, Li W, Koistinaho J. Preconditioning with spreading depression activates specifically protein kinase C δ . *Neuroreport* 2001; 12:269–273.
 61. Shamloo M, Rytter A, Wieloch T. Activation of the extracellular signal-regulated protein kinase cascade in the hippocampal CA1 region in a rat model of global cerebral ischemic preconditioning. *Neuroscience* 1999; 93:81–88.
 62. Zhao L, Nowak TS Jr. CBF changes associated with focal ischemic preconditioning in the spontaneously hypertensive rat. *J Cereb Blood Flow Metab* 2006; 26:1128–1140.
 63. Nakamura H, Katsumata T, Nishiyama Y, et al. Effect of ischemic preconditioning on cerebral blood flow after subsequent lethal ischemia in gerbils. *Life Sci* 2006; 78:1713–1719.
 64. Bastide M, Gele P, Petraut O, et al. Delayed cerebrovascular protective effect of lipopolysaccharide in parallel to brain ischemic tolerance. *J Cereb Blood Flow Metab* 2003; 23:399–405.
 65. Qin L, Wu X, Block ML, et al. Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia* 2007; 55:453–462.
 66. Eising GP, Mao L, Schmid-Schonbein GW, Engler RL, Ross J. Effects of induced tolerance to bacterial lipopolysaccharide on myocardial infarct size in rats. *Cardiovasc Res* 1996; 31:73–81.
 67. Zacharowski K, Otto M, Hafner G, et al. Endotoxin induces a second window of protection in the rat heart as determined by using p-nitro-blue tetrazolium staining, cardiac troponin T release, and histology. *Arterioscler Thromb Vasc Biol* 1999; 19:2276–2280.
 68. Colletti LM, Remick DG, Campbell DA Jr. LPS pretreatment protects from hepatic ischemia/reperfusion. *J Surg Res* 1994; 57:337–343.
 69. Fernandez ED, Flohe S, Siemens F, et al. Endotoxin tolerance protects against local hepatic ischemia/reperfusion injury in the rat. *J Endotoxin Res* 2000; 6:321–328.
 70. Heemann U, Szabo A, Hamar P, et al. Lipopolysaccharide pretreatment protects from renal ischemia/reperfusion injury: possible connection to an interleukin-6-dependent pathway. *Am J Pathol* 2000; 156:287–293.
 71. Obermaier R, Drognitz O, Grub A, et al. Endotoxin preconditioning in pancreatic ischemia/reperfusion injury. *Pancreas* 2003; 27:e51–e56.
 72. Rosenzweig HL, Minami M, Lessov NS, et al. Endotoxin preconditioning protects against the cytotoxic effects of TNF α after stroke: a novel role for TNF α in LPS-ischemic tolerance. *J Cereb Blood Flow Metab* 2007; 27:1663–1674.
 73. Barone FC, Arvin B, White RF, et al. Tumor necrosis factor- α . A mediator of focal ischemic brain injury. *Stroke* 1997; 28:1233–1244.
 74. Nawashiro H, Tasaki K, Ruetzler CA, Hallenbeck JM. TNF- α pretreatment induces protective effects against focal cerebral ischemia in mice. *J Cereb Blood Flow Metab* 1997; 17:483–490.
 75. Furuya K, Ginis I, Takeda H, et al. Cell permeable exogenous ceramide reduces infarct size in spontaneously hypertensive rats supporting in vitro studies that have implicated ceramide in induction of tolerance to ischemia. *J Cereb Blood Flow Metab* 2001; 21:226–232.
 76. Loddick SA, Turnbull AV, Rothwell NJ. Cerebral interleukin-6 is neuroprotective during permanent focal cerebral ischemia in the rat. *J Cereb Blood Flow Metab* 1998; 18:176–179.
 77. Ohtsuki T, Ruetzler CA, Tasaki K, Hallenbeck JM. Interleukin-1 mediates induction of tolerance to global ischemia in gerbil hippocampal CA1 neurons. *J Cereb Blood Flow Metab* 1996; 16:1137–1142.
 78. Bruce AJ, Boling W, Kindy MS, et al. Altered neuronal and microglial responses to excitotoxic and ischemic brain injury in mice lacking TNF receptors. *Nat Med* 1996; 2:788–794.
 79. Gary DS, Bruce-Keller AJ, Kindy MS, Mattson MP. Ischemic and excitotoxic brain injury is enhanced in mice lacking the p55 tumor necrosis factor receptor. *J Cereb Blood Flow Metab* 1998; 18:1283–1287.
 80. Spera PA, Ellison JA, Feuerstein GZ, Barone FC. IL-10 reduces rat brain injury following focal stroke. *Neurosci Lett* 1998; 251:189–192.
 81. Fan H, Cook JA. Molecular mechanisms of endotoxin tolerance. *J Endotoxin Res* 2004; 10:71–84.
 82. Garcia JH, Liu KF, Relton JK. Interleukin-1 receptor antagonist decreases the number of necrotic neurons in rats with middle cerebral artery occlusion. *Am J Pathol* 1995; 147:1477–1486.
 83. Relton JK, Martin D, Thompson RC, Russell DA. Peripheral administration of interleukin-1 receptor antagonist inhibits brain damage after focal cerebral ischemia in the rat. *Exp Neurol* 1996; 138:206–213.
 84. Raghavendra RV, Bowen KK, Dhodda VK, et al. Gene expression analysis of spontaneously hypertensive rat cerebral cortex following transient focal cerebral ischemia. *J Neurochem* 2002; 83:1072–1086.
 85. Victor NA, Wanderi EW, Gamboa J, et al. Altered PPAR γ expression and activation after transient focal ischemia in rats. *Eur J Neurosci* 2006; 24:1653–1663.
 86. Zhao Y, Patzer A, Gohlke P, et al. The intracerebral application of the PPAR γ -ligand pioglitazone confers neuroprotection against focal ischaemia in the rat brain. *Eur J Neurosci* 2005; 22:278–282.
 87. Shioda N, Ishigami T, Han F, et al. Activation of phosphatidylinositol 3-kinase/protein kinase B pathway by a vanadyl compound mediates its neuroprotective effect in mouse brain ischemia. *Neuroscience* 2007; 148:221–229.
 88. Mabuchi T, Kitagawa K, Ohtsuki T, et al. Contribution of microglia/macrophages to expansion of infarction and response of oligodendrocytes after focal cerebral ischemia in rats. *Stroke* 2000; 31:1735–1743.
 89. Lai AY, Todd KG. Microglia in cerebral ischemia: molecular actions and interactions. *Can J Physiol Pharmacol* 2006; 84:49–59.
 90. Yrjanheikki J, Keinanen R, Pellikka M, Hokfelt T, Koistinaho J. Tetracyclines inhibit microglial activation and are neuroprotective in global brain ischemia. *Proc Natl Acad Sci U S A* 1998; 95:15769–15774.
 91. Denes A, Vidyasagar R, Feng J, et al. Proliferating resident microglia after focal cerebral ischaemia in mice. *J Cereb Blood Flow Metab* 2007; 27:1941–1953.
 92. Streit WJ. Microglia as neuroprotective, immunocompetent cells of the CNS. *Glia* 2002; 40:133–139.
 93. Lalancette-Hebert M, Gowing G, Simard A, Weng YC, Kriz J. Selective ablation of proliferating microglial cells exacerbates ischemic injury in the brain. *J Neurosci* 2007; 27:2596–2605.

Correspondence: Bruce D. Trapp, PhD, Chairman, Department of Neurosciences, Lerner Research Institute, Cleveland Clinic, 9500 Euclid Avenue, NC30, Cleveland, OH 44195; trapp@ccf.org.