The Quest for Effective Treatments of Mucositis

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Commentary on "Animal Models of Mucositis: Implications for Therapy" by Joanne Bowen, Rachel Gibson, and Dorothy Keefe (page 161).

egimen-related mucosal toxicity (mucositis) is a largely unsolved medical need associated with cancer treatment. Despite its long history, patients continue to suffer and clinicians are frustrated by the diffuse, painful ulcerative lesions of the mouth, esophagus, intestine, and rectal mucosa that are commonly induced by a range of chemotherapeutics and radiation therapies and which largely defy effective intervention. This review by Bowen and colleagues largely captures the past and current roles animal modeling plays in regard to our understanding of mucositis and the quest for effective treatments.

Until the late 1990s the biological complexity of mucositis was underappreciated. Prior to that time, the condition was considered to be solely the consequence of nonspecific direct damage to rapidly dividing epithelial "stem" cells. Any thoughts of intervention were focused on palliation or modifying proliferating normal cells. Attempts to better understand the pathogenesis were largely limited to the use of cadaveric material.¹ The introduction of animal models enabled studies in which mucositis could be investigated at the tissue, cellular, and genetic levels and revealed biological targets for possible treatment.

Chemotherapy-induced mucositis has been effectively studied in the mouse, hamster, and rat. Early studies by Farrell et al,² which combined 5-fluorouracil (5-FU) and methotrexate to induce lesions of the small intestine, were important in establishing the kinetics of intestinal injury. Although largely superseded by new

hamster models, the original 5-FU-induced mucositis model in that species was critical to providing insight into the pathogenesis of mucositis.³ However, it was the advent and application of the acute radiation model in hamsters that provided the seminal data to support new mechanistic paradigms of mucositis biology at the cellular and genomic levels.^{4,5} Importantly, the results of mechanistic and genomic studies in humans corroborated the original findings in hamsters.^{6–8} The rat model introduced by Bowen and colleagues⁹ has been instrumental in further defining the complex pathoetiology of chemotherapy-induced gastrointestinal mucositis.

With the completion of phase 2 and 3 clinical trials, assessment of the translational effectiveness of animal models is now attainable. Modeling of oral and intestinal chemotherapy- or radiation-induced mucositis by Farrell et al,² Dorr et al,¹⁰ and Keefe et al¹¹ provided proof of concept for the clinical enablement of keratinocyte growth factor and confirmed information of value in establishing clinical dosing schedules. The hamster models of acute, fractionated, and chemoradiation-induced mucositis have proven to be predictive for the clinical response of at least two drugs and one biological: N-acetyl cysteine,¹² SCV-07,¹³ and FGF-20.¹⁴ In general, animal models have been most valuable in assessing potential efficacy and defining and optimizing treatment schedules.

As recognized in Keefe's dark agouti mammary adenocarcinoma model, one cannot ignore tumor response when studying potential mucositis interventions.¹¹ Before any agent can go from animal to human, it is critical that it not only shows a mucosal protection benefit but also that it fails to undermine the antitumor effects of either radiation or chemotherapy.

The candor with which Bowen and colleagues note the shortcomings of animal models of mucositis should be appreciated. Without question, both the science of mucositis and steps toward effective treatments have been furthered by the application of animal modeling. Nonetheless, Dr. Sonis is professor of Oral Medicine, Department of Oral Medicine, Infection and Immunity, Harvard School of Dental Medicine; Chief, Divisions of Oral Medicine, Brigham and Women's Hospital and the Dana-Farber Cancer Institute, Boston, Massachusetts.

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the best model is the human. Hopefully, the fruits of animal studies will continue to yield effective therapy options for patients at risk of regimen-related toxicities.

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References

PubMed ID in brackets

1. Lockhart PB, Sonis ST. Alterations in the oral mucosa caused by chemotherapeutic agents—a histologic study. J Dermatol Surg Oncol 1981;7:1019–1025.

2. Farrell CL, Bready JV, Rex KL, Chen JN, DiPalma CR, Whitcomb KL, Yin S, Hill DC, Wiemann B, Starnes CO, Havill AM, Lu ZN, Aukerman SL, Pierce GF, Thomason A, Potten CS, Ulich TR, Lacey DL. Keratinocyte growth factor protects mice from chemotherapy and radiation-induced gastrointestinal injury and mortality. Cancer Res 1998;58:933–939.

3. Sonis ST, Lindquist L, Van Vugt A, Stewart AA, Stam K, Qu GY, Iwata KK, Haley JD. Prevention of chemotherapy-induced ulcerative mucositis by transforming growth factor beta 3. Cancer Res 1999;54:1135–1138.

4. Sonis ST, Peterson RL, Edwards LJ, Lucey CA, Wang L, Mason L, Login G, Ymamkawa M, Moses G, Bouchard P, Hayes LL, Bedrosian C, Dorner AJ. Defining mechanisms of action of interleukin-11 on the progression of radiation-induced oral mucositis in hamsters. Oral Oncol 2000;36:373–381.

5. Sonis ST, Scherer J, Phelan S, Lucey CA, Barron JE, O'Donnell KE, Brennan RJ, Pan H, Busse P, Haley JD. The gene expression sequence of radiated mucosa in an animal mucositis model. Cell Prolif 2002;35(suppl 1):93–102.

6. Logan RM, Gibson RJ, Sonis ST, Keefe DM. Nuclear factor-kappaB (NF-kappaB) and cyclooxygenase-2 (COX-2) expression in the oral mucosa following cancer chemotherapy. Oral Oncol 2007;43:395–401.

7. Haddad R, Sonis S, Posner M, Wirth L, Costello R, Braschayko P, Allen A, Mahadevan A, Flynn J, Burke E, Li Y, Tishler RB. Randomized phase 2 study of concomitant chemoradiotherapy using carboplatin/paclitaxel with or without daily subcutaneous amifostine in patients with locally advanced head and neck cancer. Cancer 2009;115:4514–4523.

8. Sonis S, Haddad R, Posner M, Watkins B, Fey E, Morgan TV, Mookanamparambil L, Ramoni M. Gene expression changes in peripheral blood cells provide insight into the biological mechanisms associated with regimen-related toxicities in patients being treated for head and neck cancers. Oral Oncol 2007;43:289–300.

9. Bowen JM, Gibson RJ, Tsykin A, Stringer AM, Logan RM, Keefe DM. Gene expression analysis of multiple gastrointestinal regions reveals activation of common cell regulatory pathways following cytotoxic chemotherapy. Int J Cancer 2007;121:1847–1156.

10. Dörr W, Bässler S, Reichel S, Spekl K. Reduction of radiochemotherapy-induced early oral mucositis by recombinant human keratinocyte growth factor (palifermin): experimental studies in mice. Int J Radiat Oncol Biol Phys 2005;62:881–887.

11. Gibson RJ, Keefe DM, Clarke JM, Regester GO, Thompson FM, Goland GJ, Edwards BG, Cummins AG. The effect of keratinocyte growth factor on tumour growth and small intestinal mucositis after chemotherapy in the rat with breast cancer. Cancer Chemother Pharmacol 2002;50: 53–58.

12. Chambers MS, Welsh DV, Scrimger W, et al. RK-202 for radiationinduced oral mucositis. ASCO Annual Meeting Proceedings Part I. J Clin Oncol 2006;24:5523.

13. Watkins B, Pouliot K, Fey E, Tuthill C, Sonis S. Attenuation of radiation- and chemoradiation-induced mucositis using gamma-d-glutamyl-ltryptophan (SCV-07). Oral Dis 2010;16:655–660.

14. Alvarez E, Fey EG, Valax P, Peterson JD, Mesri M, Jeffers M, Dindinger M, Twomlow N, Ghatpande A, LaRochelle WJ, Sonis ST, Lichenstein HS. Preclinical characterization of CG53135 (FGF-20) in radiation and concomitant chemotherapy/radiation-induced oral mucositis. Clin Cancer Res 2003;9:3453–3461.