

REVIEWS

Antimicrobial De-Escalation Strategies in Hospitalized Patients With Pneumonia, Intra-Abdominal Infections, and Bacteremia

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Increasing numbers of serious hospital/healthcare- or community-acquired infections are caused by resistant (often multi-drug resistant) bacterial pathogens. Because delayed or ineffective initial therapy can have severe negative consequences, patients at risk for these types of infections typically receive initial empiric antibiotic therapy with a broad-spectrum regimen covering the most likely pathogens, based on local surveillance data and risk factors for infection with a resistant microorganism. While improving the likelihood of a successful outcome, use of broad-spectrum, often high-dose, empiric antimicrobial therapy also creates pressure for the selection or development of resistant microorganisms, as well as increasing costs and possibly exposing patients to adverse events or collateral damage such as *Clostridium difficile*-associated disease. De-escalation is a strategy that attempts to balance the competing aims of providing initial empiric therapy that is appropriate and covers the likely pathogens, and limiting antimicrobial exposure and increased

risk for emergence of resistant pathogens. More specifically, the de-escalation strategy involves collection of cultures for later microbiological assessment before initiating broad-spectrum empiric therapy covering the most likely pathogens, with the intention of streamlining or de-escalating to a more narrow-spectrum antimicrobial regimen 2–3 days later if warranted by clinical status and culture results. In some cases, negative culture results and subsequent clinical review may allow for termination of initial empiric therapy. In this manner, de-escalation enables more effective targeting of the causative pathogen(s), elimination of redundant therapy, a decrease in antimicrobial pressure for emergence of resistance, and cost savings. This article examines application of the de-escalation strategy to 3 case patients, one with healthcare-associated pneumonia, another with complicated intra-abdominal infection, and a third with central line-associated bacteremia. *Journal of Hospital Medicine* 2012;7:S13–S21. © 2012 Society of Hospital Medicine

Two conflicting aims collide when choosing initial empiric therapy for patients with a potential life-threatening infection. On the one hand, the clinical picture and seriousness of the suspected infection—sometimes with a multi-drug resistant (MDR) pathogen—point to the need for immediate empiric therapy with a broad-spectrum regimen covering the most likely pathogens. This “getting it right the first time” approach¹ is clearly a reasonable one given the significant negative impact of inappropriate or inadequate initial therapy on patient outcomes and costs,^{2–4} and the apparent inability to remedy the initial error by subsequent antimicrobial regimen adjustment.^{5–7} On the other hand, use of a broad-spectrum regimen increases the risk of emergent antimicrobial-resistant pathogens, with potential harm for the immediate patient and all subsequent patients who become exposed and infected with the resistant pathogen. Hence, the aim of optimizing initial empiric therapy

comes into conflict with an important aim of antimicrobial stewardship, namely, to use antimicrobials in a manner that does not excessively promote development or selection of antimicrobial-resistant pathogens.

The de-escalation strategy is an approach that attempts to balance these conflicting aims by providing optimal initial patient management without inordinately promoting development of antimicrobial resistance. As discussed more fully in the corresponding supplement article by Dr Syndman, the first part of this strategy involves collecting cultures from suitable patients prior to initiating broad-spectrum empiric antimicrobial therapy designed to cover the most likely pathogenic microorganisms, based on local patterns of prevalence and susceptibility, and the presence of risk factors for infection with drug-resistant species.^{8–10} The second critical step involves modification of initial empiric therapy (when warranted) based on clinical status and when culture results are available.^{8–10} In this manner, the initial broad-spectrum regimen can often be streamlined or de-escalated to a more narrow-spectrum regimen or, in some cases, terminated when negative cultures suggest no infection. Frequently, initial combination therapy can be replaced by monotherapy targeting the pathogenic organism identified in culture. Sometimes culture results indicate that initial empiric therapy was inappropriate/inadequate and requires replacement or other modification. Thus, by modifying empiric antimicrobial therapy on the basis of culture results and

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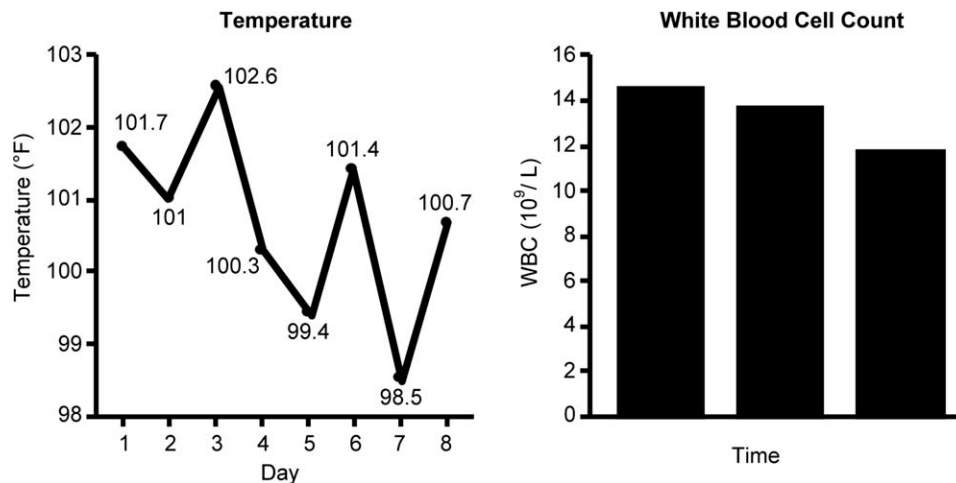


FIG. 1. Measures of body temperature and white blood cell (WBC) count for case 1 since hospital admission and initiation of empiric antibiotic therapy.

clinical criteria, the de-escalation strategy enables more effective targeting of the causative pathogen(s), elimination of redundant therapy, a decrease in antimicrobial pressure for emergence of resistance, and cost savings.^{10,11} Decreasing the number of antimicrobial agents and/or the spectrum of coverage is also expected to decrease the risk of adverse events, drug-drug interactions, and *Clostridium difficile*-associated disease.^{12,13} A number of studies have demonstrated that de-escalation of initially appropriate therapy can be successfully accomplished with either improved outcomes^{14,15} or with comparable effectiveness as continued initial therapy,^{16–18} but with reduced antimicrobial exposure and costs.¹⁹

The timing of streamlining or other modification of initial empiric therapy typically occurs when microbiological culture results become available. Assuming blood or other relevant tissue cultures were obtained prior to initiating empiric therapy, this means de-escalation or other modifications of initial therapy generally occurs 2–4 days after hospitalization and/or the beginning of empiric therapy. If rapid diagnostic tests are used to identify or rule out particular pathogens, then de-escalation may occur slightly sooner. In addition to culture results, observation of the patient in the hospital setting and improved clarity as to his or her clinical status also affect the decision about whether and how to modify the initial empiric antimicrobial regimen. The clinical scenario of the patient and his or her response to initial antimicrobial therapy is also typically clearer by day 3 of antibiotic therapy. If, for some reason, cultures were not obtained prior to beginning empiric therapy, then observations of clinical status and consideration of patient risk factors for resistant pathogens become predominant in the decision-making process. With respect to the timing of culture attainment, this should occur prior to beginning antimicrobial therapy, because therapy may reduce culture yield and result in false negative or other misleading findings.^{20,21}

CASE 1: HEALTHCARE-ASSOCIATED PNEUMONIA

Case 1 is a 72-year-old woman admitted with findings consistent with healthcare-associated pneumonia (HCAP). Empiric therapy was initiated with vancomycin and piperacillin/tazobactam. Figure 1 provides the laboratory (white blood cell [WBC] counts) and body temperature data for the patient since she entered the hospital and began empiric antibiotic therapy 3 days earlier. The WBC counts suggest the patient is responding to the antibiotic regimen, as demonstrated by a progressive reduction over the time period. However, her counts were still elevated above normal at last measurement, suggesting an incompletely resolved infection at this time. In addition, the patient is still coughing, but has less sputum production, and has some energy to get up and move around. Crackles are apparent at the right lung base. The patient's fever curve has trended down, but still shows notable fever spikes, with a temperature maximum of 101.4°F for the past 24 hours. Her blood pressure (135/84 mmHg), pulse (74 bpm), and respiratory rate (14 breaths per minute) are normal, with slightly decreased oxygen saturation (94%) on room air, although improved from initial examination 3 days earlier (92%). The blood culture shows no growth; the sputum culture simply shows oropharyngeal flora. In other words, the culture results have not isolated a causative pathogen. In addition to vancomycin and piperacillin/tazobactam, the patient continues to receive her usual medications for a past history of myocardial infarction (low-dose aspirin, metoprolol) and hypertension (enalapril, furosemide).

HCAP is a common infection often requiring initial empiric therapy with a broad-spectrum regimen that covers possible involvement of resistant bacteria. As such, HCAP frequently provides excellent opportunities for de-escalation. Figure 2 presents the general strategy from the 2005 American Thoracic Society and Infectious Diseases Society of America (ATS/IDSA)

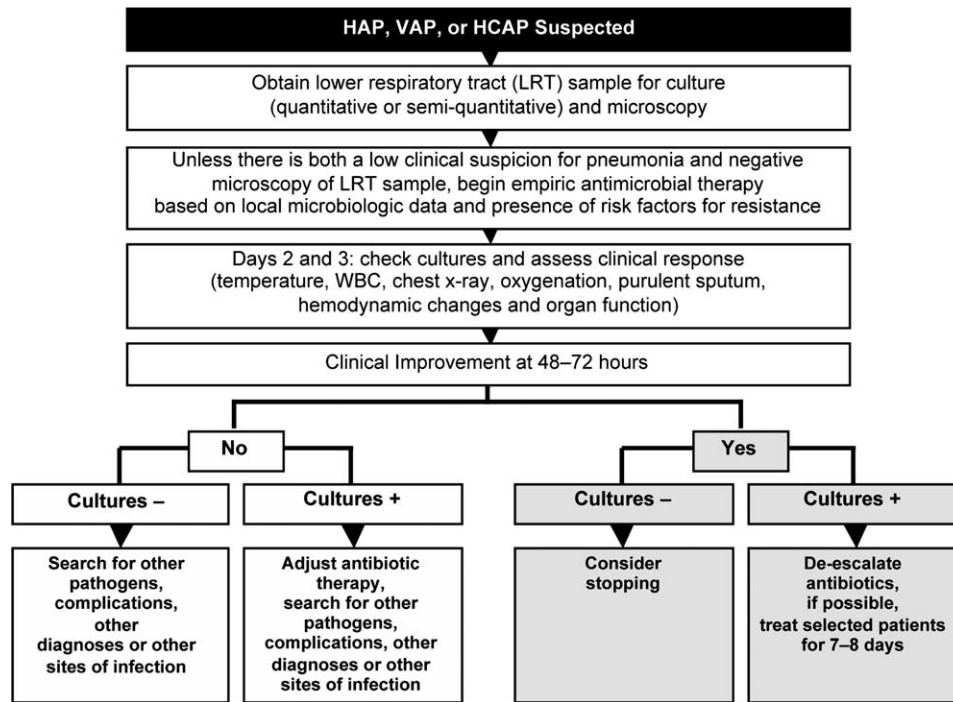


FIG. 2. Summary of management strategies for a patient with suspected hospital-acquired pneumonia (HAP), ventilator-associated pneumonia (VAP), or healthcare-associated pneumonia (HCAP). Reprinted with permission of the American Thoracic Society. Copyright© American Thoracic Society. Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. *Am J Respir Crit Care Med.* 2005;171:388–416.²² Official Journal of the American Thoracic Society. **Abbreviations:** WBC, white blood cell.

guidelines for the management of HCAP, hospital-acquired pneumonia (HAP), or ventilator-associated pneumonia (VAP).²² According to the guidelines, HCAP, HAP, and VAP should be similarly managed. Broad-spectrum initial empiric antibiotic therapy is recommended for patients with late-onset disease or those with risk factors for MDR pathogens (including high prevalence of resistance based on local antibiograms), while limited-spectrum antibiotic therapy is recommended for all other patients. Note that consideration of de-escalation or streamlining of initial therapy begins 2–3 days after initiation of therapy. Data that should be reviewed prior to instituting de-escalation include blood cultures and respiratory cultures, as well as the clinical status of the patient. The adequacy of respiratory samples used for culturing should factor into the decision-making process. For example, in patients who are not intubated or mechanically ventilated, it can be challenging to obtain a quality respiratory specimen for culture. If clinicians are uncertain as to the quality of the respiratory specimen that was cultured, then de-escalation decisions should be based more on the clinical status of the patient.

The clinical status of the patient, ≥ 2 days after beginning treatment, and culture results are critical in guiding the de-escalation process.^{9,22} The ATS/IDSA guidelines recommend serial assessments of clinical parameters to define the response to initial empiric therapy. If the therapy regimen is effective, an improvement in clinical response should be apparent within 2–3 days of its initiation.²² Hence, no change

in antimicrobial therapy should be undertaken before 3 days, unless there is evidence of rapid deterioration in clinical status or infectious diseases experts recommend a change. With respect to culture results, failure to isolate a group of MDR pathogens for which initial broad-spectrum empiric therapy was selected affords an opportunity to now streamline therapy or treat with a more narrow-spectrum regimen.⁹ Similarly, isolation of a particular pathogen can guide treatment modifications (when necessary), while a negative culture raises the possibility of terminating antimicrobial therapy, provided the culture was collected before initiating therapy. Confidence in this latter decision is bolstered when the patient exhibits rapid improvement in clinical status that is backed by radiographic resolution of lung abnormalities, or an alternative diagnosis has been established for which antimicrobial therapy is not indicated.⁹

At this stage in the process—3 days after initiating empiric therapy, and with culture results in hand and evidence of clinical improvement—the first decision or question is whether antimicrobial therapy can be stopped altogether, ie, do the current data suggest a noninfectious diagnosis (eg, pulmonary embolism, atelectasis) or that bacterial pneumonia is unlikely or has resolved. A 2000 study by Singh et al. highlighted the feasibility of using operational criteria in the form of clinical pulmonary infection score (CPIS) to decide whether to terminate or shorten the duration of initial empiric antibiotic therapy for suspected VAP.²³ More specifically, patients with pulmonary infiltrates but a

low likelihood of pneumonia (CPIS ≤ 6) were randomized to receive either standard antibiotic therapy or ciprofloxacin monotherapy. The situation was re-evaluated at 3 days, and ciprofloxacin therapy was discontinued if the CPIS remained ≤ 6 . Results showed no difference in mortality between the ciprofloxacin and standard therapy groups, despite shorter duration of therapy for the former, together with lower antimicrobial exposure and costs for the ciprofloxacin group. (Use of the CPIS to shorten the duration of empiric therapy and limit antimicrobial exposure is discussed in greater detail in the corresponding article in this supplement by Dr File.) Having said that, the case study before us describes a patient with pneumonia by clinical criteria who has responded to broad-spectrum therapy. Alternative noninfectious diagnoses are not apparent, and even though cultures have returned without significant growth, the patient should continue to receive antimicrobial treatment. The question now is whether to de-escalate/streamline to a more narrow-spectrum regimen, or continue the current one.

De-escalation often targets antimicrobials that provide unnecessarily broad coverage, eg, those with antipseudomonal activity (particularly antipseudomonal carbapenems) and/or agents with activity against methicillin-resistant *Staphylococcus aureus* (MRSA). In the absence of definitive culture results isolating a particular pathogen(s), decisions regarding which antibiotics to stop or change often depends, in large part, on patient characteristics (eg, history of prior infection with resistant pathogens, as well as drug allergies or renal insufficiency) and local antibiograms indicating the prevalence and antimicrobial susceptibility of different pneumonia pathogens in the hospital at large or particular wards within the hospital. However, negative culture results can also be useful in guiding subsequent therapy decisions or modifications. In the present case, MRSA was not grown from any cultures, and there was no evidence of Gram-positive cocci clusters with Gram staining. This suggests that vancomycin should be stopped, and antimicrobial therapy continued with a single antibiotic or antibiotic product that does include MRSA coverage. The question then is whether to continue piperacillin/tazobactam or replace it with another antibiotic.

Because *Pseudomonas aeruginosa* was not isolated, the clinician might consider streamlining piperacillin/tazobactam to an antibiotic with less pseudomonal and anaerobic coverage, possibly a nonpseudomonal third-generation cephalosporin or nonpseudomonal carbapenem, such as ertapenem. Given the activity of piperacillin/tazobactam against aerobic Gram-positive and Gram-negative pathogens, continuing piperacillin-tazobactam as single-agent therapy would also be a viable alternative. However, in the spirit of stewardship and lack of need for pseudomonal coverage, a decision was made to replace piperacillin/tazobactam

with ceftriaxone. Ceftriaxone is a nonpseudomonal third-generation cephalosporin with activity against most other Gram-negative bacteria. Note that in this case, only oropharyngeal flora grew from the respiratory culture, and the blood culture was negative. However, if a pathogen had grown from either respiratory or blood cultures, then single-agent therapy could have been used to target that specific pathogen. For example, if *Klebsiella* spp susceptible to ceftriaxone was isolated from the respiratory culture, then ceftriaxone would have been the obvious choice. If MRSA was isolated, then vancomycin (or another appropriate active agent, such as linezolid or clindamycin) could be administered as a single agent.

CASE 2: INTRA-ABDOMINAL INFECTION (DIVERTICULITIS)

Case 2 is a 56-year-old woman who presents with a diverticular abscess and walled off perforation. Interventional radiology inserts a drain, and the patient is treated with ciprofloxacin plus metronidazole. This regimen is consistent with guidelines from the Surgical Infection Society and IDSA for initial empiric treatment of complicated intra-abdominal infection of mild-to-moderate severity.²⁴ On day 3 following hospital admission and initiation of empiric therapy, the patient seems to show treatment response, as evidenced by downward trends in body temperature and WBC count (Figure 3). However, although the body temperature measures are trending in the right direction, there is still concern about continuing fever spikes and fever at last measure (100.9°F). In addition, the WBC count is still elevated, though improving. The patient's blood pressure has normalized (112/72 mmHg vs 84/58 mmHg at admission), and oxygen saturation (98%) measures are normal. The patient's lungs are clear, and her abdominal examination results are improving, though there is still some diffuse tenderness. Microbiological data show blood cultures with no growth, and isolation of Gram-negative rods from cultures of the abdominal abscess.

We now have preliminary microbiological data for a patient who remains febrile and has continuing abdominal tenderness, but who is otherwise clinically stable. Can her antimicrobial regimen be de-escalated at this point, based on what is currently known? When managing a patient after the first 3 or 4 days of empiric treatment, it is important to realize that the patient's condition with regards to infection might reflect issues unrelated to inadequate antimicrobial coverage. If the patient's clinical status has not improved, or if he or she remains febrile even 3 or 4 days into therapy, the clinician should not automatically assume the lack of improvement is due to antibiotic failure. At this point, it is important to consider possible nonantibiotic causes of persistent clinical abnormalities and fever, and for the case here, one possibility is inadequate abscess drainage. The patient

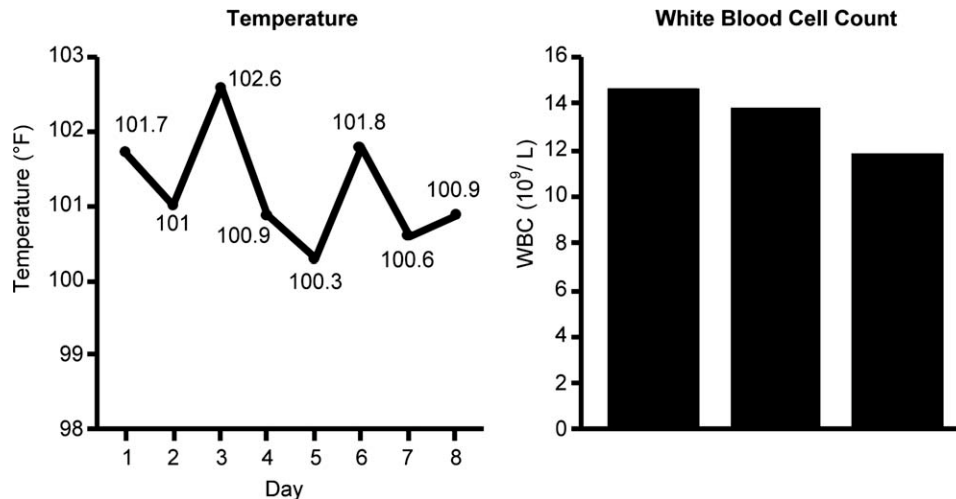


FIG. 3. Measures of body temperature and white blood cell (WBC) count for case 2 since hospital admission and initiation of empiric antibiotic therapy.

should be evaluated with abdominal imaging to ascertain whether the abscess is being adequately drained. With respect to antimicrobial therapy, the patient's blood pressure has stabilized, and her fever is trending downward. In many cases, a lingering fever such as the one observed here, in the context of improving WBC counts and clinical stabilization, may reflect inadequate mechanical drainage of the abscess. Certainly the antimicrobial therapy should not be broadened at this time, and consideration should be given to de-escalation based on the available microbiological data.

If a type of pathogenic organism is preliminarily identified from culture, but the exact identification of the organism is pending, adjustments of therapy can still be made. Adjustments can also be made based on what is *not* growing. In this case, the abscess culture has grown Gram-negative rods, but no Gram-positive organisms. Hence, continued coverage of Gram-negative organisms is warranted. In addition, anaerobes often will not readily grow in clinical cultures, and because anaerobes are frequent co-pathogens, it is appropriate to continue to provide anaerobic coverage. Based on this information, continuation of both ciprofloxacin (for aerobic Gram-negative coverage) and metronidazole (to cover for anaerobic bacteria) is appropriate in the present case. In other words, the initial empiric therapy should be continued until subsequent culture identifies a particular pathogen, at which time the therapy can be streamlined.

Now, 1 day later (day 4 of hospital admission and empiric therapy), the patient's clinical status is essentially unchanged—except for a spike in fever to 103.2°F. The WBC count is unchanged. Moreover, additional abscess culture data are available, showing definitive identification of an extended-spectrum β -lactamase (ESBL)-producing *Escherichia coli* organism. The blood culture is still negative. The first observation is that ESBL-producing *E coli* is a relatively unusual pathogen in a community-based infection.

However, the patient here did have risk factors for antibiotic-resistant pathogens, notably prior antimicrobial therapy as an outpatient. It is also important to recognize that community-acquired infections with ESBL-producing bacteria (mostly isolated from the urinary tract) have been reported in many parts of the world, and even in some parts of the United States.²⁵

Based on these additional microbiological data, the patient was switched to treatment with ertapenem, a nonpseudomonal carbapenem with activity against ESBL-producing Enterobacteriaceae.²⁶ In addition, ertapenem, and other carbapenems, have excellent activity against anaerobes,²⁶ and it is prudent to continue coverage for anaerobes even though anaerobes were not grown in the culture. As mentioned above, these organisms are difficult to grow in clinical culture, and they are common pathogens or co-pathogens in intra-abdominal infections. Carbapenems are widely regarded as the antimicrobials of choice for treatment of serious, invasive infections with ESBL-producing bacteria.²⁷ Furthermore, by choosing a nonpseudomonal carbapenem, compared with an antipseudomonal carbapenem, the new antibiotic regimen provides coverage of the isolated ESBL-producing *E coli* organism—as well as covering possible anaerobe involvement—without exposing host bacteria to unnecessarily broad antipseudomonal activity. Cephalosporins, monobactams, and fluoroquinolones are generally not active against ESBL-producing Enterobacteriaceae, and β -lactam/ β -lactamase inhibitor combinations (eg, ampicillin/sulbactam, piperacillin/tazobactam) do not have reliable activity in serious, high inoculum infections caused by ESBL-producing Enterobacteriaceae.²⁷

CASE 3: CENTRAL LINE-ASSOCIATED BLOODSTREAM INFECTION

Case 3 is a 56-year-old man who presented to the hospital emergency department with status epilepticus. He was intubated, had a central line placed in the

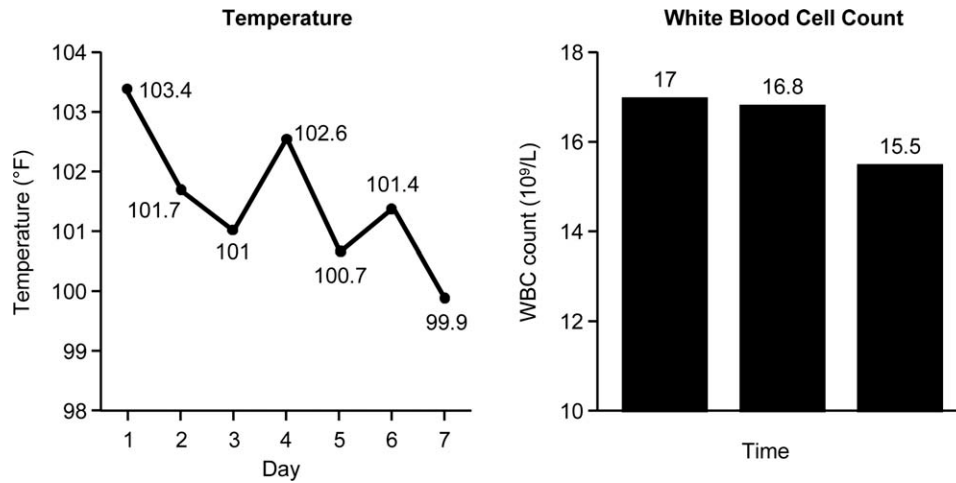


FIG. 4. Measures of body temperature and white blood cell (WBC) count for case 3 after 7 days in the intensive care unit (ICU), and after narrowing the initial combination antibiotic regimen to vancomycin monotherapy on day 6.

internal jugular vein, and was admitted to the intensive care unit (ICU). The seizure was successfully broken by aggressive treatment with repeated intravenous dosing of lorazepam and loading with fosphenytoin. Empiric antibiotic therapy was initiated with vancomycin and piperacillin/tazobactam on day 5, after spiking a fever of 103.4°F. No clear source of the fever was identified. While in the ICU with a central line in place, 2 sets of blood cultures were drawn. Now on hospital day 6, the patient is still spiking fever, although the fever trend appears to be decreasing. The patient is hemodynamically stable, with no other abnormal findings (besides persistent fever) on physical examination. WBC count remains elevated, and both sets of blood cultures are notable for growth of Gram-positive cocci.

Bloodstream infection is a serious condition in hospitalized patients that is associated with significant morbidity and mortality.²⁸ Patients with suspected bloodstream infection typically receive empiric broad-spectrum antimicrobial therapy, and are thus good candidates for de-escalation based on subsequent clinical status and blood culture results. Because of the seriousness of bloodstream infection, healthcare workers are sometimes hesitant to de-escalate initial empiric therapy, even when cultures isolate a pathogen susceptible to narrower-spectrum agents, particularly if the patient appears to be improving on such therapy. This is true for various serious hospital or healthcare-associated infections,^{16,29} but particularly for bloodstream infections. Moreover, when central line-associated bloodstream infection (CLABSI) is suspected, the most important initial intervention is to remove the infected central venous catheter. For a patient with a short-term catheter and a CLABSI due to Gram-negative bacilli, *S aureus* (which appears to be a likely pathogen for the case patient here), enterococci, fungi, or mycobacteria, the 2009 IDSA guidelines for management of intravascular catheter-related

infections recommend catheter removal.³⁰ Catheter removal is even more important than antibiotic coverage; this point cannot be stressed enough. In some extreme cases, when the line cannot be removed for clinical reasons, antibiotic lock therapy can be used to supplement systemic antimicrobial therapy.³⁰ This involves instilling a high antibiotic solution into the catheter lumen for a period of time in order to sterilize the lumen and prevent biofilm formation.³¹

The first step taken for the patient here was to remove the central venous catheter. Then, turning to the preliminary culture data, there is evidence for Gram-positive cocci in the patient's blood. The blood culture did not grow any Gram-negative organisms. Gram-positive cocci (coagulase-negative staphylococci, *S aureus* [methicillin-susceptible or MRSA]) are the most common causes of CLABSI.³² Can the physician de-escalate antibiotic therapy in this patient with CLABSI based on the preliminary information? Yes. The information is solid enough to suggest removal of the catheter which was providing coverage for Gram-negative bacteria (piperacillin/tazobactam), while continuing vancomycin for coverage of possible MRSA, pending further review, ie, until the Gram-positive cocci are speciated. Rapid diagnostic methods, including polymerase chain reaction (PCR) and nucleic acid probes, can be used to provide more information about certain pathogens (such as MRSA^{33,34}) before final culture and susceptibility results are available, but these are not routinely available in many clinical microbiology laboratories. Furthermore, these newer technologies remain fairly expensive.

Revisiting the patient 1 day later (hospital day 7), after narrowing the initial combination antibiotic regimen to vancomycin monotherapy, the physical examination indicates the patient is clinically stable, with continued improvement in fever and WBC count (Figure 4). Blood culture analysis now isolates methicillin-susceptible *S aureus* (MSSA). Methicillin

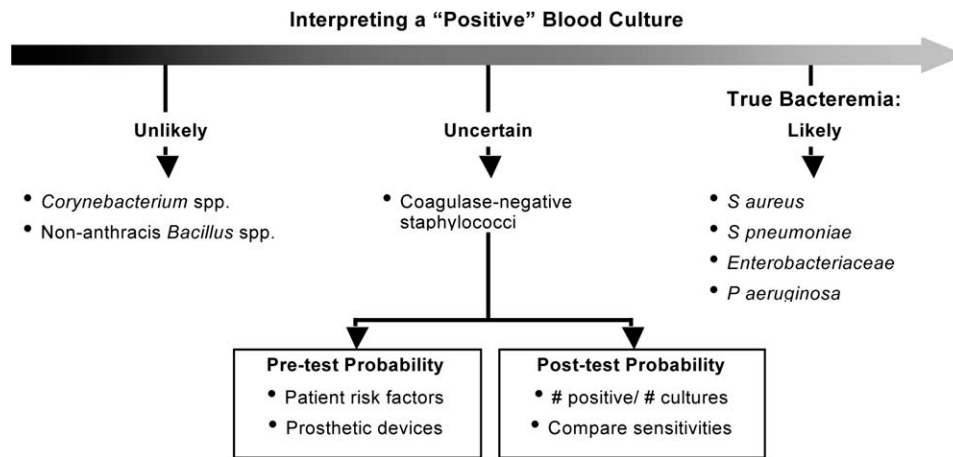


FIG. 5. Schematic of process using species and strain identification to determine the significance of coagulase-negative staphylococci (CoNS) isolated from blood cultures. (Data, in part, based on study by Kim et al.⁴²)

resistance mediates resistance to all β -lactams, including carbapenems, greatly limiting treatment options. Vancomycin is the most commonly utilized antibiotic for the treatment of MRSA, and the recent clinical practice guidelines from the IDSA recommend either vancomycin or daptomycin for management of MRSA bacteremia in adult patients.³⁵ However, anti-staphylococcal penicillins and first-generation cephalosporins are the antibiotics of choice for MSSA infections, and particularly for MSSA bloodstream infections.

The activity provided by vancomycin (or daptomycin) is overly broad if MSSA is involved, and importantly, it is not as effective as treatment with an antistaphylococcal penicillin or first-generation cephalosporin. A recent study by Stryjewski et al., of hemodialysis patients with MSSA bacteremia, reported a higher proportion of treatment failure with vancomycin versus first-generation cephalosporin therapy (31% vs 13%; $P = 0.02$).³⁶ Furthermore, multivariate analysis identified vancomycin (vs first-generation cephalosporin) use as a significant independent predictor of treatment failure (odds ratio [OR], 3.53; 95% confidence interval [CI], 1.15–13.45; $P = 0.04$). Similarly, Chang et al. reported nafcillin, an antistaphylococcal penicillin, was superior to vancomycin in preventing bacteriologic failure (persistent failure and/or relapse) in patients with MSSA bacteremia (0% vs 19%; $P = 0.058$), and used multivariate analysis to identify vancomycin as a significant independent predictor of relapse (OR, 6.5; 95% CI, 1.0–52.8; $P < 0.05$).³⁷ Another recent study by Lodise et al. reported that initial empiric therapy with vancomycin for endocarditis caused by MSSA was associated with a higher infection-related mortality rate than initial empiric therapy with a β -lactam-containing regimen (39% vs 11%; $P = 0.005$).³⁸ The negative impact of initial treatment with vancomycin persisted even in patients switched to a β -lactam therapy after culture results became available.

Hence, if a patient is being treated with vancomycin for a bloodstream (or other) infection due to MSSA, the therapy is suboptimal. In such a scenario—which corresponds to that for the case patient here—vancomycin should be discontinued and replaced with an antistaphylococcal penicillin or first-generation cephalosporin. Many times, clinicians are resistant to terminating vancomycin and de-escalating to antistaphylococcal penicillin/first-generation cephalosporin therapy in a patient with bacteremia who is apparently responding to vancomycin. However, as the studies just reviewed make clear, not only is vancomycin treatment overly broad for the circumstance, it is also suboptimal and does not represent best clinical practice or patient care. Furthermore, continuing vancomycin in this situation unnecessarily exposes the patient to possible renal toxicity, particularly when aggressive dosing or prolonged vancomycin treatment is involved.³⁹ Because of these issues and concerns, case 3 was de-escalated from vancomycin to cefazolin, a first-generation cephalosporin. One word of caution, however, is that there is some controversy over using cefazolin in patients with *S aureus* native valve endocarditis, given the possibility of a Type A β -lactamase-producing species causing cefazolin degradation.⁴⁰ As a result, the clinician should first rule out endocarditis in the patient here before proceeding with cefazolin therapy. Another alternative would be to use an antistaphylococcal penicillin, such as nafcillin.

Finally, when dealing with bacteremia, and particularly when dealing with a possible CLABSI, the issue of potential culture contamination needs to be seriously considered and answered. Treating an actual infection, not what appears to be an infection because of culture contamination, is particularly important when dealing with possible CLABSI, because coagulase-negative staphylococci (CoNS) are the most common cause of these types of infections,³² and CoNS are also frequent blood-culture contaminants.⁴¹ Therefore, one needs to determine whether a blood

culture growing a CoNS represents true bacteremia or simply contamination—which will obviously impact de-escalation decisions.

In addition, when determining whether a blood culture is truly “positive” and clinically significant, it is important to consider whether the isolated pathogens are unlikely to be contaminants, likely to be contaminants, or the situation is unclear. A 2000 study by Kim et al.⁴² suggested that, among patients with ≥ 2 positive blood cultures for CoNS, routine identification of CoNS species and genotyping selected isolates using pulsed-field gel electrophoresis may improve the process of discriminating contaminants from pathogens. Various additional factors need to be weighed when trying to interpret CoNS blood culture results, including patient risk factors, presence of prosthetic devices, number of blood cultures and number positive, and the antimicrobial sensitivity patterns of different isolates. For example, if the sensitivity patterns of 2 CoNS strains isolated from a patient are the same, the likelihood is increased that they represent true pathogens rather than contaminants. Figure 5 presents a schematic of this general approach.⁴²

CONCLUSIONS

De-escalation is a critical component of antimicrobial stewardship. As the prevalence of antimicrobial resistance grows in the hospital and community, de-escalation will have an increasingly important role in limiting the further emergence of antimicrobial resistance. Pneumonia, intra-abdominal infection, and bloodstream infection are commonly managed in the hospital setting. Each of these infection types presents excellent opportunities for de-escalation, and each presents unique challenges and caveats. Concerted efforts must be made by clinicians and stewardship personnel to de-escalate as soon as possible, based on culture results and clinical status. Although not discussed here, successful de-escalation programs utilize structured process, guidelines, and algorithms to consistently implement de-escalation efforts. These tools of implementation are more fully discussed in the corresponding article in this supplement by Dr Rosenberg.

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References

- Kollef M. Appropriate empirical antibacterial therapy for nosocomial infections: getting it right the first time. *Drugs*. 2003;63:2157–2168.
- Garnacho-Montero J, Ortiz-Leyba C, Herrera-Melero I, et al. Mortality and morbidity attributable to inadequate empirical antimicrobial therapy in patients admitted to the ICU with sepsis: a matched cohort study. *J Antimicrob Chemother*. 2008;61:436–441.
- Kollef KE, Schramm GE, Wills AR, Reichley RM, Micek ST, Kollef MH. Predictors of 30-day mortality and hospital costs in patients with ventilator-associated pneumonia attributed to potentially antibiotic-resistant gram-negative bacteria. *Chest*. 2008;134:281–287.
- Kollef MH, Sherman G, Ward S, Fraser VJ. Inadequate antimicrobial treatment of infections: a risk factor for hospital mortality among critically ill patients. *Chest*. 1999;115:462–474.
- Luna CM, Vujacich P, Niederman MS, et al. Impact of BAL data on the therapy and outcome of ventilator-associated pneumonia. *Chest*. 1997;111:676–685.
- Montravers P, Gauzit R, Muller C, Marmuse JP, Fichelle A, Desmonts JM. Emergence of antibiotic-resistant bacteria in cases of peritonitis after intraabdominal surgery affects the efficacy of empirical antimicrobial therapy. *Clin Infect Dis*. 1996;23:486–494.
- Zilberberg MD, Shorr AF, Micek ST, Mody SH, Kollef MH. Antimicrobial therapy escalation and hospital mortality among patients with health-care-associated pneumonia: a single-center experience. *Chest*. 2008;134:963–968.
- Lisboa T, Rello J. De-escalation in lower respiratory tract infections. *Curr Opin Pulm Med*. 2006;12:364–368.
- Niederman MS. The importance of de-escalating antimicrobial therapy in patients with ventilator-associated pneumonia. *Semin Respir Crit Care Med*. 2006;27:45–50.
- Paterson DL. Impact of antibiotic resistance in gram-negative bacilli on empirical and definitive antibiotic therapy. *Clin Infect Dis*. 2008;47(suppl 1):S14–S20.
- Dellit TH, Owens RC, McGowan JE Jr, et al. Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America guidelines for developing an institutional program to enhance antimicrobial stewardship. *Clin Infect Dis*. 2007;44:159–177.
- Bad Bugs, No Drugs: As Antibiotic R&D Stagnates, a Public Health Crisis Brews. Alexandria, VA: Infectious Diseases Society of America, 2004.
- Spellberg B, Guidos R, Gilbert D, et al. The epidemic of antibiotic-resistant infections: a call to action for the medical community from the Infectious Diseases Society of America. *Clin Infect Dis*. 2008;46:155–164.
- Giantsou E, Liratzopoulos N, Efraimidou E, et al. De-escalation therapy rates are significantly higher by bronchoalveolar lavage than by tracheal aspirate. *Intensive Care Med*. 2007;33:1533–1540.
- Kollef MH, Morrow LE, Niederman MS, et al. Clinical characteristics and treatment patterns among patients with ventilator-associated pneumonia. *Chest*. 2006;129:1210–1218.
- Alvarez-Lerma F, Alvarez B, Luque P, et al. Empiric broad-spectrum antibiotic therapy of nosocomial pneumonia in the intensive care unit: a prospective observational study. *Crit Care*. 2006;10:R78.
- Eachempati SR, Hydo LJ, Shou J, Barie PS. Does de-escalation of antibiotic therapy for ventilator-associated pneumonia affect the likelihood of recurrent pneumonia or mortality in critically ill surgical patients? *J Trauma*. 2009;66:1343–1348.
- Leone M, Garcin F, Bouvenot J, et al. Ventilator-associated pneumonia: breaking the vicious circle of antibiotic overuse. *Crit Care Med*. 2007;35:379–385; quiz 386.
- Berild D, Mohseni A, Diep LM, Jensenius M, Ringertz SH. Adjustment of antibiotic treatment according to the results of blood cultures leads to decreased antibiotic use and costs. *J Antimicrob Chemother*. 2006;57:326–330.
- Hummel M, Warga C, Hof H, Hehlmann R, Buchheidt D. Diagnostic yield of blood cultures from antibiotic-naïve and antibiotic-treated patients with haematological malignancies and high-risk neutropenia. *Scand J Infect Dis*. 2009;41:650–655.
- Souweine B, Veber B, Bedos JP, et al. Diagnostic accuracy of protected specimen brush and bronchoalveolar lavage in nosocomial pneumonia: impact of previous antimicrobial treatments. *Crit Care Med*. 1998;26:236–244.
- Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. *Am J Respir Crit Care Med*. 2005;171:388–416.
- Singh N, Rogers P, Atwood CW, Wagener MM, Yu VL. Short-course empiric antibiotic therapy for patients with pulmonary infiltrates in the intensive care unit. A proposed solution for indiscriminate antibiotic prescription. *Am J Respir Crit Care Med*. 2000;162:505–511.
- Solomkin JS, Mazuski JE, Bradley JS, et al. Diagnosis and management of complicated intra-abdominal infection in adults and children: guidelines by the Surgical Infection Society and the Infectious Diseases Society of America. *Clin Infect Dis*. 2010;50:133–164.
- Peirano G, Costello M, Pitout JD. Molecular characteristics of extended-spectrum beta-lactamase-producing *Escherichia coli* from the Chicago area: high prevalence of ST131 producing CTX-M-15 in community hospitals. *Int J Antimicrob Agents*. 2010;36:19–23.
- Keating GM, Perry CM. Ertapenem: a review of its use in the treatment of bacterial infections. *Drugs*. 2005;65:2151–2178.
- Pitout JD. Infections with extended-spectrum beta-lactamase-producing Enterobacteriaceae: changing epidemiology and drug treatment choices. *Drugs*. 2010;70:313–333.
- Valles J, Ferrer R. Bloodstream infection in the ICU. *Infect Dis Clin North Am*. 2009;23:557–569.
- Drew RH, White R, MacDougall C, Hermesen ED, Owens RC Jr. Insights from the Society of Infectious Diseases Pharmacists on antimicrobial stewardship guidelines from the Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America. *Pharmacotherapy*. 2009;29:593–607.

30. Mermel LA, Allon M, Bouza E, et al. Clinical practice guidelines for the diagnosis and management of intravascular catheter-related infection: 2009 Update by the Infectious Diseases Society of America. *Clin Infect Dis*. 2009;49:1–45.
31. Yahav D, Rozen-Zvi B, Gaftor-Gvili A, Leibovici L, Gaftor U, Paul M. Antimicrobial lock solutions for the prevention of infections associated with intravascular catheters in patients undergoing hemodialysis: systematic review and meta-analysis of randomized, controlled trials. *Clin Infect Dis*. 2008;47:83–93.
32. Leonidou L, Gogos CA. Catheter-related bloodstream infections: catheter management according to pathogen. *Int J Antimicrob Agents*. 2010;36(suppl 2):S26–S32.
33. Hombach M, Pfyffer GE, Roos M, Lucke K. Detection of methicillin-resistant *Staphylococcus aureus* (MRSA) in specimens from various body sites: performance characteristics of the BD GeneOhm MRSA assay, the Xpert MRSA assay, and broth-enriched culture in an area with a low prevalence of MRSA infections. *J Clin Microbiol*. 2010;48:3882–3887.
34. Stamper PD, Cai M, Howard T, Speser S, Carroll KC. Clinical validation of the molecular BD GeneOhm StaphSR assay for direct detection of *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* in positive blood cultures. *J Clin Microbiol*. 2007;45:2191–2196.
35. Liu C, Bayer A, Cosgrove SE, et al. Clinical practice guidelines by the Infectious Diseases Society of America for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children. *Clin Infect Dis*. 2011;52:e18–e55.
36. Stryjewski ME, Szczech LA, Benjamin DK Jr, et al. Use of vancomycin or first-generation cephalosporins for the treatment of hemodialysis-dependent patients with methicillin-susceptible *Staphylococcus aureus* bacteremia. *Clin Infect Dis*. 2007;44:190–196.
37. Chang FY, Peacock JE Jr, Musher DM, et al. *Staphylococcus aureus* bacteremia: recurrence and the impact of antibiotic treatment in a prospective multicenter study. *Medicine (Baltimore)*. 2003;82:333–339.
38. Lodise TP Jr, McKinnon PS, Levine DP, Rybak MJ. Impact of empirical-therapy selection on outcomes of intravenous drug users with infective endocarditis caused by methicillin-susceptible *Staphylococcus aureus*. *Antimicrob Agents Chemother*. 2007;51:3731–3733.
39. Jeffres MN, Isakow W, Doherty JA, Micek ST, Kollef MH. A retrospective analysis of possible renal toxicity associated with vancomycin in patients with health care-associated methicillin-resistant *Staphylococcus aureus* pneumonia. *Clin Ther*. 2007;29:1107–1115.
40. Nannini EC, Singh KV, Murray BE. Relapse of type A beta-lactamase-producing *Staphylococcus aureus* native valve endocarditis during cefazolin therapy: revisiting the issue. *Clin Infect Dis*. 2003;37:1194–1198.
41. Viagappan M, Kelsey MC. The origin of coagulase-negative staphylococci isolated from blood cultures. *J Hosp Infect*. 1995;30:217–223.
42. Kim SD, McDonald LC, Jarvis WR, et al. Determining the significance of coagulase-negative staphylococci isolated from blood cultures at a community hospital: a role for species and strain identification. *Infect Control Hosp Epidemiol*. 2000;21:213–217.